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•Original article•

Synthesis and anti-HIV activities of phorbol derivatives

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[ABSTRACT] In this study, 37 derivatives of phorbol esters were synthesized and their anti-HIV-1 activities evaluated, building upon our previous synthesis of 51 phorbol derivatives. 12-Para-electron-acceptor-*trans*-cinnamoyl-13-decanoyl phorbol derivatives stood out, demonstrating remarkable anti-HIV-1 activities and inhibitory effects on syncytia formation. These derivatives exhibited a higher safety index compared with the positive control drug. Among them, 12-(*trans*-4-fluorocinnamoyl)-13-decanoyl phorbol, designated as compound 3c, exhibited the most potent anti-HIV-1 activity (EC₅₀ 2.9 nmol·L⁻¹, CC₅₀/EC₅₀ 11 117.24) and significantly inhibited the formation of syncytium (EC₅₀ 7.0 nmol·L⁻¹, CC₅₀/EC₅₀ 4891.43). Moreover, compound 3c is hypothesized to act both as an HIV-1 entry inhibitor and as an HIV-1 reverse transcriptase inhibitor. Isothermal titration calorimetry and molecular docking studies indicated that compound 3c may also function as a natural activator of protein kinase C (PKC). Therefore, compound 3c emerges as a potential candidate for developing new anti-HIV drugs.

[KEY WORDS] Phorbol esters; Anti-HIV-1 activity; Syncytia formation; 12-(*Trans*-4-fluorocinnamoyl)-13-decanoyl phorbol; Safety index; HIV-1 entry inhibitor; HIV-1 reverse transcriptase inhibitor; PKC activator

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Introduction

The relentless spread of acquired immunodeficiency syndrome (AIDS) globally has catalyzed the extensive development of anti-HIV medications, targeting the eradication of causative agents ^[1]. In this quest, a multitude of plant-derived compounds are being scrutinized for their potential to inhibit HIV-1 replication and its crucial enzymes. Central to this exploration are phorbol esters, tetracyclic diterpenoids predominantly found in the *Euphorbiaceae* and *Thymelaeaceae* fam-

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ilies [2-6]. These esters are known for their diverse biological activities, including inhibition of HIV-1 protease, activation of protein kinase C (PKC), platelet aggregation, cell differentiation, anti-tuberculosis properties, cytotoxic activities, and the activation of mesenchymal stem cells [3-5, 7]. Interestingly, some phorbol esters, such as 12-O-tetradecanoyl phorbol 13-acetate (TPA), are known for their tumor-promoting and pro-inflammatory properties and are used to induce inflammation in pharmacological and biochemical research. In contrast, compounds like 12-deoxyphorbol-13-acetate (prostratin) and phorbol 12-acetate-13-decanoate have shown nontumor-promoting activities, along with pronounced anti-HIV effects [4, 6-8]. Moreover, phorbol 12-acetate-13-decanoate and its 12-benzoyl derivative have demonstrated notable anti-HIV activities [9-11]. The compound 4-deoxy-4 β -phorbol-12-tiglate-13-phenylacetate, effective in reactivating 70%-75% of latent HIV at concentrations ranging from 9.1–0.091 μmol L⁻¹, has been identified as a promising lead for the development of anti-HIV drugs targeting latency reactivation [5].

Extracting phorbol esters from plants is fraught with challenges such as labor-intensive processes, difficult separations, and low yields. Consequently, the focus has shifted to



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These authors have no conflict of interest to declare.

the structural modification of phorbol, a field that has reached maturity, using methods such as selective hydroxyl group protection, esterification, and deprotection. Notably, the hydroxyl groups in phorbol exhibit a reactivity order of C20-OH > C13-OH > C12-OH > C4-OH > C9-OH [6], guiding the structural modifications primarily towards the C20, C13 and C12 hydroxyl groups. Previous research primarily explored the esterification of phorbol with long-chain saturated fatty acids [6].

The concept of privileged substructures and pharmacophore merging is pivotal in drug design and the structural modification of natural products ^[9, 12, 13]. In this study, 35 phorbol esters derivatives, including phorbol monoesters, diesters and triesters, 4α-phorbol esters, and 4α-4-deoxyphorbol esters were designed and synthesized by coupling with cinnamic acid derivatives, oleanolic acid, glycyrrhetinic acid, unsaturated and saturated fatty acids, and terminal brominated saturated fatty acids (Figs. 1 and 2). This study also encompassed the evaluation of the anti-HIV-1 activities of these derivatives, along with 51 previously synthesized phorbol ester derivatives ^[14, 15].

Results and Discussion

Synthesis of phorbol esters

In this research, we synthesized an array of 35 phorbol

ester derivatives, including variants of 4α -phorbol ester and 4α -4-deoxyphorbol ester. The initial step in the synthesis involved protecting the C20-OH group of phorbol with trityl chloride, followed by esterification using cinnamic acid, its derivatives, or other carboxylic acids in different proportions. The subsequent removal of the trityl group with 3% HClO₄/ MeOH resulted in the production of 12,13-phorbol diester derivatives (1a-1k, 3a-3g and 4a-4b) and a 13-phorbol ester (6c). In a parallel synthesis route, phorbol was acylated with acetic anhydride and subsequently esterified with carboxylic acids. The C-20-O-acetyl or C-13,20-O-diacetyl groups were then removed using either 3% HClO₄/MeOH or 0.05 mol·L⁻ KOH/MeOH, yielding 12,13-phorbol diester derivatives (2a-**2h**) and a 13-phorbol ester (5a). The synthesis of the 4α -phorbol ester and 4α -4-deoxyphorbol ester (10, 11) mirrored the approach used for compounds 1a-1k. The synthetic pathways for these phorbol derivatives (1a-1k, 2a-2h, 3a-3g, 4a-4b, 5a, 6c, 10 and 11) are detailed in Schemes 1-5.

For the synthesis of the target compounds 4c, 6a-6b, and 9a-9b containing dual pharmacophores, we employed methods outlined in Schemes 6-7. The process began with the esterification of phorbol-13,20-diacetate with 6-bromohexanoic acid, followed by coupling with uracil to yield intermediate A and the target compound 9a. To create intermediate B_1-B_3 , 20-O-trityl-phorbol was first esterified with acetic

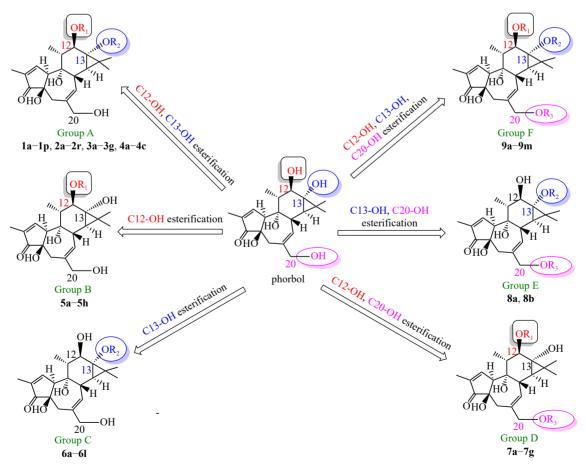


Fig. 1 The classifications of phorbol derivitives.

Scheme 1 Synthesis of compounds **1a–1k** reagents and conditions: (a) Ph₃CCl, pyridine, 3 d; (b) 3 eq RCOOH, DMAP, EDCI, DCM, overnight; (c) 3% HClO₄/MeOH, 60 min.

Scheme 2 Synthesis of compounds **3a–3g** reagents and conditions: (a) Ph₃CCl, pyridine, 3 d; (c) 3% HClO₄/MeOH, 60 min. (d) 1.5 eq RCOOH, DMAP, EDCI, DCM, overnight; (e) 1.5 eq decanoic acid, DMAP, EDCI, DCM overnight.

Phorbol Phorbol-13,20-diacetate (3) Phorbol-13,20-diacetate (3) Phorbol-13,20-diacetate (3) Phorbol-13,20-diacetate (3)
$$OR_1$$
 OR_2 OR_3 OR_4 OR_5 OR_5 OR_5 OR_5 OR_5 OR_5 OR_6 OR_6

Scheme 3 Synthesis of compounds **2a-2h**, **5a** reagents and conditions: (d) 1.5 eq RCOOH, DMAP, EDCI, DCM, overnight; (f) acetic anhydride, Et₃N, DCM, overnight; (g) 3% HClO₄/MeOH, overnight; (h) 0.05 mol·L⁻¹ KOH/MeOH, 30 min.

anhydride and either 6-bromohexanoic acid or 9-bromononanoic acid. After removing the C-20-O-acetyl group using 3% HClO₄/MeOH, the intermediates were coupled with uracil or piperazine. Furthermore, oleanolic acid and glycyrrhetic acid were reacted with 1,4-dibromobutane to produce intermediate C, which was subsequently coupled with intermediate A or B_1 – B_3 to synthesize the target compounds 9b, 4c, and 6a–6b.

Anti-HIV-1 activities of phorbol esters

The anti-HIV-1 activities of 88 phorbol ester derivatives, including 51 phorbol esters derivatives synthesized by our team, were evaluated [14, 15]. As shown in Table 1, 12,13-didecanoyl-phorbol (**1b**, EC₅₀ 4.6 nmol·L⁻¹) exhibited a much higher anti-HIV-1 activity than 12,13-didecanoyl- 4α -phorbol (**10**, EC₅₀ 122 510 nmol·L⁻¹) and 12,13-didecanoyl- 4α -4-

deoxyphorbol (11, EC₅₀ 3763.3 nmol·L⁻¹), highlighting the critical influence of the 4 β -hydroxyl group. As shown in Tables 1–4, esterification with the same acid leads to varying levels of anti-HIV-1 activity, depending on the number of ester groups in the phorbol molecule. Phorbol-12,13,20-triesters generally show significantly lower activity than monoesters and diesters, such as phorbol-12,13,20-tricinnamate (9c, EC₅₀ > 200 000 nmol·L⁻¹), phorbol-12,20-dicinnamate (7e, EC₅₀ 27 010.0 nmol·L⁻¹), phorbol-12-cinnamate (5d, EC₅₀ 19 785.0 nmol·L⁻¹), phorbol-13-cinnamate (6f, EC₅₀ 6655.0 nmol·L⁻¹), phorbol-12,13-dicinnamate (1l, EC₅₀ 123.6 nmol·L⁻¹), and phorbol-12,13-diacetate (9f, EC₅₀ 323.5 nmol·L⁻¹). In a striking finding presented in Table 4, phorbol esters with C13-OH and C20-OH of phorbol esterified by the same acid but with C12-

Scheme 4 Synthesis of compounds **4a-4b**, **6c** reagents and conditions: (a) Ph₃CCl, pyridine, 3 d; (c) 3% HClO₄/MeOH, 60 min. (d) 1.5 eq RCOOH, DMAP, EDCI, DCM, overnight.

Scheme 5 Synthesis of compounds **10** and **11** reagents and conditions: (a) Ph₃CCl, pyridine, 3 d; (b) 3 eq RCOOH, DMAP, EDCI, DCM, overnight; (c) 3% HClO₄/MeOH, 60 min.

OH modified by a different acid exhibited highly effective anti-HIV activities, such as 12-eicosapentaenoyl phorbol-13, 20-diacetate ($\bf{9j}$, EC₅₀ 70.5 nmol·L⁻¹) and 12-*trans*-cinnamoyl phorbol-13,20-dibutyryl esters ($\bf{9d}$, EC₅₀ 54.9 nmol·L⁻¹). In contrast, compounds coupled with oleanolic acid ($\bf{9b}$) and phorbol itself ($\bf{9a}$) showed minimal activity (EC₅₀ > 200 000 nmol·L⁻¹). Furthermore, the 12-*trans*-cinnamoyl-13-dibutyryl-20-(3,4-methylenedioxycinnamoyl)-phorbol esters ($\bf{9l}$), synthesized with different acids, also demonstrated a significant anti-HIV-1 activity (EC₅₀ 671.4 nmol·L⁻¹).

As shown in Tables 1, 2 and 4, phorbol esters coupled with oleanolic acid (9b) and phorbol itself (9a) showed no anti-HIV-1 activity. Conversely, those coupled with gly-cyrrhetic acids (4c, 6a, and 6b) exhibited weak anti-HIV activity (EC $_{50}$: 6a > 6b > 4c). Compounds 6a and 6b were phorbol-13-monoester. Compound 4c was 12-acetyl-phorbol-13-ester. It is speculated that the anti-HIV-1 activity is influenced by the number of methylene groups in the linker arm and the ester group at the C12 position, especially when phorbol is combined with large pharmacophores like glycyrrhetic acid.

As shown in Table 2, phorbol was esterified with the same acid to produce phorbol-12-monoesters or phorbol-13-monoesters. Phorbol-13-monoesters consistently demonstrated stronger anti-HIV-1 activities than their 12-monoester counterparts. For example, 13-eicosanyol phorbol (6k) had an

EC₅₀ of 20.2 nmol·L⁻¹, notably lower than 12-eicosanyol phorbol (5c, EC₅₀ 39 955.0 nmol·L⁻¹), and 13-eicosapentaenoyl phorbol (6i) exhibited a notably lower EC50 than 12eicosapentaenoyl phorbol (5g) (136.2 nmol·L⁻¹ vs 5325.0 nmol·L⁻¹). This trend was consistent, with the exception of biotinylated derivatives (5a, 6c, $EC_{50} > 200~000~\text{nmol}\cdot\text{L}^{-1}$). As shown in Tables 1 and 3, the anti-HIV-1 activity of phorbol-13,20-diesters or phorbol-12,20-diester was much lower than that of phorbol-12,13-diester. For instance, 13,20-dieicosapentaenoyl phorbol (8b, EC₅₀ of 12 740.0 nmol·L⁻¹) and 12-eicosapentaenoyl phorbol-20-acetate (7d, EC₅₀ 5080.0 nmol·L⁻¹) were markedly less potent than 12-eicosapentaenoyl phorbol-13-acetate (2m, EC_{50} 4.1 nmol·L⁻¹). Phorbol was esterified by the same saturated fatty acid, yielding phorbol-12,13-diester. As shown in Table 1, the anti-HIV-1 activity of phorbol-12,13-diester did not increase with the increase in methylene number. 12,13-Didecanoyl phorbol exhibited the strongest anti-HIV-1 activity (1b, EC₅₀ 4.6 nmol·L⁻¹) with a high safety index (CC₅₀/EC₅₀ 23 665.22); 12,13-di (3-phenylpropanoyl)-phorbol showed the stronger anti-HIV-1 activity (1m, EC₅₀ 26.4 nmol·L⁻¹) than phorbol-12,13-diester obtained by combining phorbol with unsaturated acid; 12,13-di (6-bromohexanoyl)-phorbol exhibited high anti-HIV-1 activity (1f, EC₅₀ 5.7 nmol·L⁻¹). As shown in Table 1, the anti-HIV-1 activity of phorbol-12,13-diester obtained by combining phorbol with terminal brominated satur-

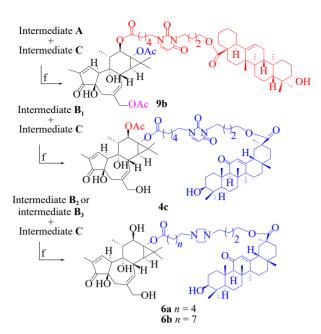
Scheme 6 Synthesis of compound **9a** reagents and conditions: (a) 6-bromohexanoic acid or 9-bromononanoic acid, DMAP, EDCI, DCM overnight; (b) uracil or piperazine, K₂CO₃, DMF, 60 °C or 50 °C, 12 h; (c) acetic anhydride, Et₃N, DCM, overnight; (d) 3% HClO₄/MeOH, 60 min; (e) 1,4-dibromobutane, K₂CO₃, DMF, 12 h.

ated fatty acids decreased with the increase in methylene number. For example, 12,13-di (9-bromononanoyl)-phorbol (1h) had a lower EC₅₀ value of 17 413.2 nmol·L⁻¹ than 12,13di (10-bromodecanoyl)-phorbol (1i, EC_{50} 19 873.3 nmol·L⁻¹). As shown in Table 1, most 13-acetyl-phorbol-12-esters exhibited significant anti-HIV-1 activities, such as 12-docosahexaenoyl-phorbol-13-acetate (2n, EC_{50} 8.3 nmol·L⁻¹). However, 12-(6-bromohexanoyl)-13-acetyl-phorbol did not show anti-HIV-1 activity (2a, $EC_{50} > 200\ 000\ nmol \cdot L^{-1}$). As shown in Table 1, the anti-HIV-1 activities of compounds 2b and 2c increased with the increase in methylene number; the anti-HIV-1 activities of 13-acetyl-phorbol-12-trans-cinnamovl derivatives decreased with the introduction of para electron-acceptor groups in the cinnamoyl moiety. For instance, 12-transcinnamoyl phorbol-13-acetate (2i) exhibited a dramatically lower EC₅₀ value than 12-feruloyl-13-acetyl-phorbol (2j) $(14.0 \text{ nmol} \cdot \text{L}^{-1} \text{ vs } 77 \text{ } 920.0 \text{ nmol} \cdot \text{L}^{-1})$. As shown in Table 1, 13-butyryl phorbol-12-unsaturated acid derivatives showed high anti-HIV-1 activity, such as 12-trans-cinnamoyl-13-butyryl phorbol (20, EC₅₀ 7.9 nmol·L⁻¹). As shown in Table 1, a series of 13-decanoyl-phorbol-12-trans-cinnamoyl derivatives all exhibited remarkable anti-HIV-1 activities with a

high safety index. Moreover, the anti-HIV activity could be increased by incorporating the *para* electron-acceptor groups into the cinnamoyl group ($3\mathbf{a}-3\mathbf{e}$), so the anti-HIV-1 activities of these derivatives were stronger than that of positive control drug (AZT, EC₅₀ 5.4 nmol·L⁻¹). Among these derivatives, 12-(*trans*-4-fluorocinnamoyl)-13-decanoyl phorbol emerged as the most potent compound ($3\mathbf{c}$, EC₅₀ 2.9 nmol·L⁻¹) with a high safety index (CC₅₀/EC₅₀ 11 117.24), but its anti-HIV-1 activity decreased after 13-decanoyl was replaced by 6-bromohexanoyl ($3\mathbf{f}$, EC₅₀ 301.9 nmol·L⁻¹), as shown in Table 1.

Inhibition of syncytia formation induced by HIV-1_{IIIR}

The data in Table 5 illustrated that compounds **1b**, **2m**, **2n**, **2q**, and **3a–3d**, recognized for their remarkable anti-HIV-1 activities, also demonstrated significant inhibition of syncytia formation induced by HIV-1_{IIIB}. In particular, compounds **1b** and **3a–3d** outperformed the positive control drug 3TC in terms of inhibition strength and safety (**1b**, EC₅₀ 5.2 nmol·L⁻¹, CC₅₀/EC₅₀ 20 550.00; **3a**, EC₅₀ 26.2 nmol·L⁻¹, CC₅₀/EC₅₀ 1438.36; **3b**, EC₅₀ 9.6 nmol·L⁻¹, CC₅₀/EC₅₀ 3368.23; **3c**, EC₅₀ 7.0 nmol·L⁻¹, CC₅₀/EC₅₀ 4891.43; **3d**, EC₅₀ 33.7 nmol·L⁻¹, CC₅₀/EC₅₀ 1564.24; 3TC, EC₅₀ 175.8



Scheme 7 Synthesis of compounds **6a**, **6b**, **9b**, and **4c** reagents and conditions: (f) K₂CO₃, KI, MeCN or DMF, 75 °C, 36 h.

nmol·L⁻¹, $CC_{50}/EC_{50} > 1137.66$).

A comprehensive analysis of anti-HIV-1 activities and syncytia formation inhibition highlighted the superior performance of 13-decanoyl-phorbol-12-*trans*-cinnamoyl derivatives, especially those with *para* electron-acceptor groups in the cinnamoyl structure. The introduction of *para* electron-withdrawing groups, such as fluorine (F), chlorine (Cl), or bromine (Br), into the 12-*trans*-cinnamoyl phorbol derivatives (compounds 3c, 3e, and 3b) significantly enhanced their anti-HIV-1 activities compared with derivatives with electron-donating groups like -OCH₃ or -OCH₂O- (compounds 3d and 3a). There is a clear trend indicating that the stronger the electron absorption capacity of the substituents (ranked as -F > -Cl > -Br), the greater the anti-HIV-1 activity.

Potential of compound 3c as an inhibitor of HIV-1 $_{\rm IIIB}$ replication at the entry step

We investigated the mechanism by which compound 3c inhibits HIV-1 replication, particularly examining its impact on HIV-1 reverse transcription and virus entry processes. To understand the role of compound 3c in HIV-1 replication, we analyzed its effect on the intermediates during reverse transcription, specifically single-stranded DNA (ssDNA) and 2-long terminal repeat (2LTR) circles. The results demonstrated that compound 3c significantly reduced the expression of these intermediate products, indicating its inhibitory role in HIV-1 replication either during the HIV-1 reverse transcription stage or in preceding steps (Fig. 3).

To further determine whether compound 3c functions as an inhibitor at the virus entry step, we treated C8166 cells with this compound at two critical points: during the infection by HIV-1 $_{\rm IIIB}$ and after the virus had entered the cells. The results revealed that the treatment with compound 3c during

the HIV-1_{IIIB} infection phase effectively suppressed virus replication. However, its inhibitory effect was notably reduced after the virus had entered the cells. These observations suggest that compound **3c** likely functions as an HIV-1 entry inhibitor, in a manner similar to T-20, a known inhibitor of this stage (Fig. 4).

Effects of compound 3c on PKC

We then explored the role of compound 3c as a potential natural activator of PKC, inspired by its structural similarity to diacylglycerol (C3-C=O, C4-β-OH, and C20-OH). This similarity suggests that phorbol esters, such as compound 3c, might activate PKC. Furthermore, the specific substitutions at C-12 and C-13 in the phorbol esters are believed to influence their interaction with PKC's hydrophobic region, affecting both the depth and orientation of their insertion into the enzyme [16]. A molecular docking study was conducted to analyze the interaction between compound 3c and PKC. As depicted in Fig. 5, compound 3c exhibited high binding energy $(-8.32 \text{ kcal} \cdot \text{moL}^{-1})$ at the active site of PKC- δ . The interactions between compound 3c and PKC-δ primarily involved the formation of hydrogen bonds with GLN-25 and hydrophobic interactions with other amino acid residues, such as PHE-27, ALA-23 and, ASN-24. The binding affinity of compound 3c for PKC-C1 protein was further quantified using isothermal titration calorimetry (ITC). The results revealed that compound 3c exhibits a high affinity for PKC, with a dissociation constant (K_d) of 10.0 ± 1.3 nmol·L⁻¹. This finding indicates that compound 3c may act as a natural activator of PKC.

Stability of compound 3c in plasma in vitro

The stability of compound **3c** in plasma *in vitro* was investigated. Specifically, the concentration of compound **3c** in rat plasma over time was monitored using high-performance liquid chromatography (HPLC). As shown in Fig. 6, the findings indicated that compound **3c** maintained its stability in rat plasma *in vitro* throughout the duration of the experiment. Notably, there was no significant degradation of the compound, nor was there any metabolization back to its parent compound, phorbol.

Experimental

General experimental procedures

Reagents and solvents were of commercial quality and used without further purification. ¹H and ¹³C NMR spectra were obtained from Varian UNITY INOVA-600 (Agilent, USA) with tetramethylsilane (TMS) as an internal standard. High-resolution mass spectra of synthesized compounds were recorded on Agilent 6540 UHD Q-TOF (Agilent, USA) with an electrospray ionization (ESI) interface. Thin-layer chromatography (TLC) analysis was performed on Huanghai silica gel GF-254 plates (Yantai Huanghai, 02025892, Y.K.X. Material Technology Co., Ltd., China). Compounds were purified by silica gel column (Yantai Huanghai, AKX-H0090, Y.K.X. Material Technology Co., Ltd., China). The structures of all the phorbol esters (1a–9b), 4α-phorbol ester (10)

Table 1 Anti-HIV-1 activity and safety index of 12,13-phorbol diesters.

Na	$\mathbf{R_{i}}$	$ m R_2$	D	Anti-HIV activity	Cytotoxicity	Safety index
No.	N ₁	K ₂	R ₃	(EC ₅₀ , nmol·L ⁻¹)	(CC ₅₀ , μmol·L ⁻¹)	(CC_{50}/EC_{50})
1a	Pelargonyl	Pelargonyl	Н	$35\ 270 \pm 18215.1$	22.7291 ± 2.3457	/
1b	Decanoyl	Decanoyl	Н	4.6 ± 1.1	106.865 ± 13.9371	23 665.22
1c	Undecanoyl	Undecanoyl	Н	3180 ± 381.8	> 200	> 62.89
1d	Dodecanoyl	Dodecanoyl	Н	16510 ± 1866.8	> 200	> 19.60
1e	Myristoyl	Myristoyl	Н	18670 ± 2775.5	> 200	> 12.12
1f	6-Bromohexanoyl	6-Bromohexanoyl	Н	5.7 ± 4.1	22.205 ± 1.1384	3895.61
1g	7-Bromoheptanoyl	7-Bromoheptanoyl	Н	180 ± 8.1	79.71 ± 7.3822	4428.33
1h	9-Bromononanoyl	9-Bromononanoyl	Н	17130 ± 400.5	74.0345 ± 10.9126	> 11.63
1i	10-Bromodecanoyl	10-Bromodecanoyl	Н	19873.3 ± 5354.7	26.1641 ± 2.7342	/
1j	11-Bromoundecanoyl	11-Bromoundecanoyl	Н	$80\ 850 \pm 17\ 740$	> 200	> 2.96
1k	Biotin	Biotin	Н	> 200 000	> 200	/
11	trans-Cinnamoyl	trans-Cinnamoyl	Н	123.6 ± 73.5	26.145 ± 10.7268	211.53
1m	3-Phenylpropanoyl	3-Phenylpropanoyl	Н	26.4 ± 10.5	26.34 ± 4.7659	997.73
1n	trans-2-Butenoyl	trans-2-Butenoyl	Н	622.4 ± 29.1	77.565 ± 15.9877	124.62
10	Eicosapentaenoyl	Eicosapentaenoyl	Н	1280.6 ± 409.3	> 200	> 156.18
1p	Eicosanoyl	Eicosanoyl	Н	$87\ 425 \pm 1732.4$	> 200	> 2.29
2a	6-Bromohexanoyl	Ac	Н	> 200 000	> 200	/
2 b	7-Bromoheptanoyl	Ac	Н	47.4 ± 37.3	73.6 ± 2.6446	1554.47
2c	8-Bromooctanoyl	Ac	Н	33.5 ± 8.1	80.565 ± 1.0253	2441.21
2d	3,4-Methylene dioxycinnamoyl	Ac	Н	22.5 ± 15.3	104.845 ± 9.7934	4659.78
2e	4-Bromocinnamoyl	Ac	Н	18.8 ± 0.8	41.15 ± 9.1358	2188.83
2f	4-Fluorocinnamoyl	Ac	Н	26.4 ± 1.1	107.125 ± 8.2237	4057.76
2g	4-Methoxycinnamoyl	Ac	Н	47 ± 20.1	108.575 ± 9.5106	2310.11
2h	Biotin	Ac	Н	> 200 000	> 200	> 11.26
2i	trans-Cinnamoyl	Ac	Н	14 ± 9.5	75.055 ± 20.4425	5361.07
2 j	Feruloyl	Ac	Н	$77\ 920 \pm 18\ 144.4$	97.47 ± 9.9985	1.25
2k	4-Chlorocinnamoyl	Ac	Н	24.6 ± 8.3	23.17 ± 4.2992	945.71
21	Ac	Ac	Н	323.6 ± 116.6	> 200	> 618.24
2m	Eicosapentaenoyl	Ac	Н	4.1 ± 0.8	26.405 ± 11.2076	6440.24
2n	Docosahexaenoyl	Ac	Н	8.3 ± 4.0	44.12 ± 2.5173	5315.66
20	trans-Cinnamoyl	Butyryl	Н	7.9 ± 2.4	32.335 ± 9.8924	4093.04
2p	Feruloyl	Butyryl	Н	95.7 ± 22.5	77.84 ± 0.9192	813.38
2q	Eicosapentaenoyl	Butyryl	Н	7.4 ± 2.8	23.265 ± 3.8537	3165.31
2r	Docosahexaenoyl	Butyryl	Н	28.3 ± 11.2	26.7 ± 2.6304	946.99
3a	3,4-Methylene dioxycinnamoyl	Decanoyl	Н	4.8 ± 2.0	37.685 ± 1.5627	7851.04
3b	4-Bromocinnamoyl	Decanoyl	Н	3.8 ± 0.5	32.335 ± 8.0539	8841.43
3c	4-Fluorocinnamoyl	Decanoyl	Н	2.9 ± 0.8	34.24 ± 1.7819	11 117.24
3d	4-Methoxycinnamoyl	Decanoyl	Н	4.5 ± 0.0013	52.715 ± 7.3893	11 714.44
3e	4-Chlorocinnamoyl	Decanoyl	Н	3.4 ± 0.1	33.485 ± 8.2519	9848.53
3f	trans-Cinnamoyl	Decanoyl	Н	301.9 ± 61.5	128.8774 ± 57.872	562.49
3g	Biotin	Decanoyl	Н	17260 ± 354.2	17.1095 ± 1.0309	/
4a	Decanoyl	Biotin	Н	18545 ± 120.2	16.3950 ± 0.5673	/
4b	4-Fluorocinnamoyl	6-Bromohexanoyl	Н	153.62 ± 41.3	48.1037 ± 28.6126	444.86
4c	Ac	A	Н	$75\ 870 \pm 33\ 474.4$	> 200	> 2.01
10	Decanoyl	Decanoyl	Н	122510 ± 6912.7	> 200	1.63
11	Decanoyl	Decanoyl	Н	6230 ± 3488.4	59.5516 ± 2.5928	/
AZT				5.4 ± 3.0	> 200	> 26 666.60

Compounds 1l–1p and 2i–2r were phorbol esters previously synthesized by our team [14, 15], compound 10 was 4α-phorbol derivative; compound 11 was 4α -4-deoxyphorbol derivative; A: 3-(4-butyl glycyrrhetinic acid)-1-(6-oxohexyl) pyrimidine-2,4(1H,3H)-dione; AZT: zidovudine.



Table 2 Anti-HIV-1 activity and safety index of 12-phorbol esters and 13-phorbol esters.

No.	R ₁	\mathbf{R}_2	R ₃	Anti-HIV activity (EC ₅₀ , nmol·L ⁻¹)	Cytotoxicity (CC ₅₀ , μmol·L ⁻¹)	Safety index (CC ₅₀ /EC ₅₀)
5a	Biotin	Н	Н	> 200 000	> 200	/
5b	Docosahexaenoyl	Н	Н	5060 ± 1060.7	46.3 ± 0.5657	9.15
5c	Eicosanoyl	Н	Н	$39\ 955 \pm 18\ 151.4$	> 200	> 5.01
5d	trans-Cinnamoyl	Н	Н	$19\ 785 \pm 120.2$	114.055 ± 17.8403	5.76
5e	3-Phenylpropanoyl	Н	Н	$51\ 575 \pm 6936.7$	> 200	> 3.88
5f	trans-2-Butenoyl	Н	Н	> 200 000	> 200	/
5g	Eicosapentaenoyl	Н	Н	5325 ± 473.8	44.795 ± 17.5433	8.41
5h	Myristoyl	Н	Н	1625 ± 544.5	23.885 ± 1.7324	14.70
6a	Н	В	Н	$98\ 200 \pm 14\ 241.1$	16.3522 ± 0.4254	/
6b	Н	C	Н	$96~005 \pm 4278$	18.9147 ± 0.1387	/
6c	Н	Biotin	Н	> 200 000	> 200	/
6d	Н	Docosahexaenoyl	Н	95.1 ± 27.4	46.865 ± 0.4596	492.80
6e	Н	3-Methoxy-4-acet oxycinnamoyl	Н	9540 ± 2404.2	92.23 ± 8.556	9.67
6f	Н	trans-Cinnamoyl	Н	6655 ± 1958.7	> 200	> 30.65
6g	Н	3-Phenylpropanoyl	Н	$11\ 140 \pm 7028.6$	> 200	> 17.95
6h	Н	trans-2-Butenoyl	Н	$21\ 990 \pm 3139.6$	> 200	> 9.09
6i	Н	Eicosapentaenoyl	Н	136.2 ± 52.3	72.985 ± 18.2363	535.87
6j	Н	Angeloyl	Н	$44\ 515 \pm 20\ 810.2$	> 200	> 4.49
6k	Н	Eicosanoyl	Н	41.7 ± 8.7	90.195 ± 14.1351	4478.41
61	Н	Decanoyl	Н	563.5 ± 196.3	117.56 ± 7.9196	208.62
AZT				5.4 ± 3.0	> 200	> 26 666.60

Compounds **5b–5h** and **6d–6l** were phorbol esters previously synthesized by our team [14,15]; B: 1-(4-butyl glycyrrhetinic acid)-4-(6-oxohexyl)-piperazine; C: 1-(4-butyl glycyrrhetinic acid)-4-(9-oxononyl)- piperazine; AZT: zidovudine.

Table 3 Anti-HIV-1 activity and safety index of 12,20-phorbol diesters and 13,20-phorbol diesters.

No.	R ₁	R ₂	R ₃	Anti-HIV activity (EC ₅₀ , nmol·L ⁻¹)	Cytotoxicity (CC ₅₀ , µmol·L ⁻¹)	Safety index (CC ₅₀ /EC ₅₀)
7a	Myristoyl	Н	Ac	4415 ± 459.6	25.26 ± 6.5195	> 5.72
7b	trans-Cinnamoyl	Н	Ac	$29\ 420\pm 1117.2$	108.39 ± 22.3304	3.68
7c	3-Phenylpropanoyl	Н	Ac	$46\ 345 \pm 22\ 478.9$	118.8 ± 1.5415	2.56
7d	Eicosapentaenoyl	Н	Ac	5080 ± 84.9	33.91 ± 9.2207	6.68
7e	trans-Cinnamoyl	Н	trans-Cinnamoyl	$27\ 010 \pm 16\ 136.2$	> 200	> 7.41
7 f	Eicosanoyl	Н	Ac	> 200 000	> 200	/
7 g	Docosahexaenoyl	Н	Ac	$15\ 660 \pm 2220.3$	55.9 ± 12.5865	3.57
8a	Н	trans-2-Butenoyl	trans-2-Butenoyl	$64\ 790 \pm 17\ 903.9$	144.25 ± 13.1098	2.23
8b	Н	Eicosapentaenoyl	Eicosapentaenoyl	$12\ 740 \pm 9772.2$	86.42 ± 16.7584	6.78
AZT				5.4 ± 3.0	> 200	> 26 666.60

Compounds 7a-7g and 8a-8b were phorbol esters previously synthesized by our team [14,15]; AZT: zidovudine.

and 4α -4-deoxyphorbol ester (11) were characterized through 1D (1 H and 13 C) NMR and HR-ESI-MS data analysis. Following literature methods $^{[17]}$, 20-O-trityl-phorbol (intermedi-

ate compound 1, 70%) was synthesized from phorbol (isolated and identified by our laboratory, HPLC purity: 96.61%). From this compound, 13-decanoyl-20-*O*-trityl-phorbol (inter-



Table 4 Anti-HIV-1 activity and safety index of phorbol-12,13,20-triesters.

No.	$\mathbf{R_{i}}$	\mathbf{R}_2	R_3	Anti-HIV activity (EC ₅₀ , nmol·L ⁻¹)	Cytotoxicity (CC ₅₀ , μ mol·L ⁻¹)	Safety index (CC ₅₀ /EC ₅₀)
9a	D	Ac	Ac	> 200 000	> 200	/
9b	Е	Ac	Ac	> 200 000	> 200	/
9c	trans-Cinnamoyl	trans-Cinnamoyl	trans-Cinnamoyl	> 200 000	> 200	/
9d	trans-Cinnamoyl	Butyryl	Butyryl	55 ± 11.7	> 200	> 3639.67
9e	3-Methoxy-4-acet oxycinnamoyl	Butyryl	Butyryl	293.2 ±186.3	> 200	> 682.24
9f	Ac	Ac	Ac	8955 ± 770.7	> 200	> 22.33
9g	Eicosapentaenoyl	Eicosapentaenoyl	Eicosapentaenoyl	41 740 ± 24 621.5	129.78 ± 23.278	3.12
9h	3-Phenylpropanoyl	Ac	Ac	208.7 ± 53.5	125.08 ± 12.7986	599.47
9i	trans-2-Butenoyl	Ac	Ac	2180 ± 989.9	> 200	> 91.74
9j	Eicosapentaenoyl	Ac	Ac	70.5 ± 45.6	54.22 ± 7.2832	1015.99
9k	trans-Cinnamoyl	Butyryl	trans-Cinnamoyl	469.6 ± 11.2	> 200	> 425.94
91	trans-Cinnamoyl	Butyryl	3,4-Methylene dioxycinnamoyl	671.4 ± 205.6	> 200	> 297.89
9m	3,4-Methylene dioxycinnamoyl	Ac	Ac	210.1 ± 44.9	> 200	> 952.15
AZT				5.4 ± 3.0	> 200	> 26 666.60

Compounds **9c–9m** were phorbol esters previously synthesized by our team ^[14, 15]; D: 3-[6-(12-*O*-hexanoyl phorbol-13,20-diethyl ester)]-1-(6-oxohexyl) pyrimidine-2,4(1*H*,3*H*)-dione; E: 3-(4-butyl oleanolic acid)-1-(6-oxohexyl) pyrimidine-2,4(1*H*,3*H*)-dione; AZT: zidovudine.

Table 5 Inhibition of syncytia formation induced by HIV-1_{IIIB} and safety index of phorbol derivatives.

No.	R1	R2	R3	Anti-HIV activity (EC ₅₀ , nmol·L ⁻¹)	Cytotoxicity (CC ₅₀ , µmol·L ⁻¹)	Safety index (CC ₅₀ /EC ₅₀)
1b	Decanoyl	Decanoyl	Н	5.2 ± 1.4	106.865 ± 13.9371	20 550.00
2m	Eicosapentaenoyl	Ac	Н	60.6 ± 9.6	26.405 ± 11.2076	435.73
2n	Docosahexaenoyl	Ac	Н	943.1 ± 63.4	44.120 ± 2.5173	46.78
2q	Eicosapentaenoyl	Butyryl	Н	$11\ 66.7 \pm 76.1$	23.265 ± 3.8537	19.94
3a	3,4-Methyl enedi oxycinnamoyl	Decanoyl	Н	26.2 ± 5.1	37.685 ± 1.5627	1438.36
3b	4-Bromocinnamoyl	Decanoyl	Н	9.6 ± 5.2	32.335 ± 8.0539	3368.23
3c	4-Fluorocinnamoyl	Decanoyl	Н	7.0 ± 2.1	34.240 ± 1.7819	4891.43
3d	4-Methoxycinnamoyl	Decanoyl	Н	33.7 ± 2.3	52.715 ± 7.3893	1564.24
3TC				175.8 ± 9.2	> 200	> 1137.66

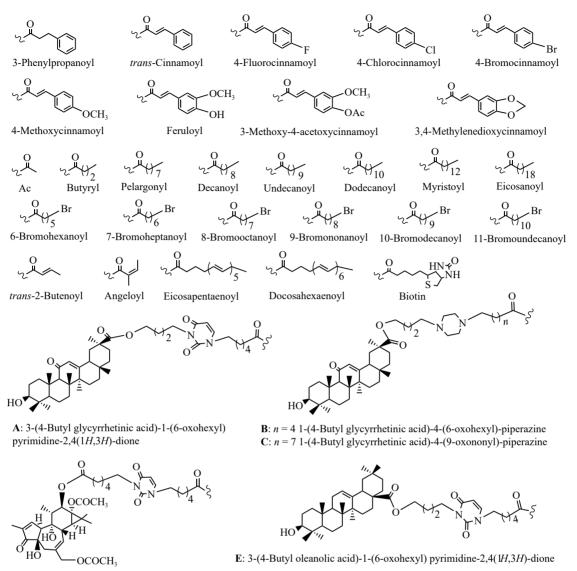
Compounds 2m, 2n, and 2q were phorbol esters derivatives previously synthesized by our team [14, 15]; 3TC: lamivudine.

mediate compound 2) was obtained with an 80% yield. Additionally, phorbol-13,20-diacetate (intermediate compound 3) was synthesized from phorbol, achieving a 78% yield (Schemes 1–3). The detailed procedures for the synthesis of intermediate compounds 1–3 are available in the Supporting Information.

Synthesis of target compounds 1a-1k, 2a-2h, 3a-3g, 4a-4b, 5a, 6c, 10, and 11

A solution of 20-*O*-trityl-phorbol (1, 0.5 mmol) in 12 mL dichloromethane (DCM) was treated with various acids (non-anoic acid, decanoic acid, hendecoic acid, dodecanoic acid, myristic acid, 6-bromohexanoic acid, 7-bromoheptanoic acid, 9-bromononanoic acid, 10-bromodecanoic acid, 11-bromo-

hendecoic acid, and biotin) in 3.0 equialents (eq). 1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDCI) (1.5 eq) and *N*,*N*-dimethyl-4-aminopyridine (DMAP) (1.0 eq) were added. Chemicals and reagents were purchased from Adamas, HPLC purity 97%–99%, Titan Technology Co., Ltd. (Shanghai, China). After overnight stirring, the reaction was quenched with 20 mL of saturated NaHCO₃, and then the mixture was extracted three times with DCM. The combined organic layers were concentrated under reduced pressure and further purified by silica gel column chromatography (petroleum ether: ethyl acetate, 5:1) to yield phorbol-12,13,20-triesters. These triesters were then treated with 10 mL of 3% HClO₄/MeOH. After stirring for 1 h, the mixture was neutral-



D: 3-(6-(12-*O*-Hexanoyl phorbol-13,20-diethyl ester))-1-(6-oxohexyl) pyrimidine-2,4(1*H*,3*H*)-dione

Fig. 2 The acyl groups of phorbol esters.

ized with 1 mol·L⁻¹ NaOH and then extracted with DCM. The organic layers were concentrated under reduced pressure. The final products were purified by silica gel column chromatography (petroleum ether: ethyl acetate, 3:1 to 1:1) to afford compounds 1a (80%), 1b (75%), 1c (83%), 1d (75%), 1e (81%), 1f (75%), 1g (65%), 1h (88%), 1i (90%), 1j (86%) as colorless oil, and 1k (60%) as white amorphous powder, respectively (Scheme 1).

The procedures for the synthesis of target compounds 2a-2h and 3a-3g were similar to those for 1a-1k. Phorbol-12,13,20-triesters were synthesized by esterification of phorbol-13,20-diacetate (3, 0.5 mmol) with various acids (6-bromohexanoic acid, 7-bromoheptanoic acid, 8-bromooctanoic acid, 3,4-methylenedioxycinnamic acid, 4-bromocinnamic acid, 4-fluorocinnamic acid, 4-methoxycinnamic acid, and biotin) in 1.5 eq. The triesters were treated with 3% HClO₄/MeOH for 12 h. Subsequently, the products were isolated and

purified by silica gel chromatography and eluted with (petroleum ether: ethyl acetate, 2:1) to afford **2a** (80%), **2b** (83%), **2c** (85%), **2d** (77%), **2e** (82%), **2f** (76%), **2g** (88%), and **2h** (38%) as white amorphous powders (Scheme 3). Phorbol-12,13,20-triesters were synthesized by esterification of phorbol-13,20-diacetate (**3**, 0.5 mmol) and biotin (1.5 eq). Later, these triesters were treated with 0.05 mol·L⁻¹ KOH/MeOH for 30 min to obtain compound **5a** (60%) as a white amorphous powder (Scheme 3).

20-*O*-Trityl-13-decanoyl-phorbol (**2**, 0.5 mmol) was reacted separately with 3,4-methylenedioxycinnamic acid, 4-bromocinnamic acid, 4-fluorocinnamic acid, 4-methoxycinnamic acid, 4-chlorcinnamic acid, cinnamic acid, and biotin (1.5 eq) to yield compounds **3a**–**3g**. Reaction mixtures were eluted with petroleum ether and ethyl acetate (5 : 1) by silica gel chromatography to yield **3a** (70%), **3b** (86%), **3c** (90%), **3d** (83%), **3e** (88%), **3f** (86%), and **3g** (48%) as white

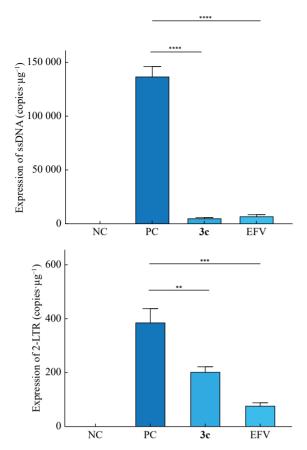


Fig. 3 Effects of compound 3c on the intermediate products of HIV-1 reverse transcription. ssDNA: single-stranded DNA; 2LTR: 2 long terminal repeat; EFV: efavirenz, HIV-1 reverse transcriptase inhibitor, used as a approved drug control; NC: negative control, only C8166 cells, used as a negative control; PC: positive control, only virus infected C8166 cells, without drug treatment, serve as a positive control. Data are presented as means \pm SD of three independent experiments. **P < 0.01, ****P < 0.001, ****P < 0.0001 vs positive control group.

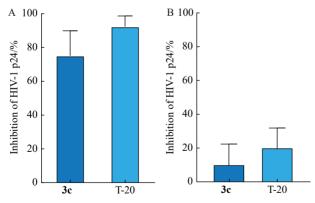


Fig. 4 Effects of compound 3c after the virus had entered the cells. (A) When HIV-1_{IIIB} entered C8166 cells, the cells were treated with compound 3c; (B) After the HIV-1_{IIIB} entered C8166 cells, the cells were treated with compound 3c. T-20: enfuvirtide, HIV-1 entry inhibitor. Data are presented as means \pm SD of three independent experiments.

amorphous powders (Scheme 2).

As shown in Scheme 4, compound 6c (65%, white amorphous powder) was synthesized by esterification of 20-*O*-trityl-phorbol (1, 0.5 mmol) and biotin (1.5 eq); compound 4a (35%, a white amorphous powder) was synthesized by esterification of 20-*O*-trityl-phorbol (1, 0.5 mmol), biotin (1.5 eq), and decanoic acid (1.5 eq); compound 4b (55%, a white amorphous powder) was synthesized similarly by using 20-*O*-trityl-phorbol (1, 0.5 mmol) and 6-bromohexanoic acid (1.5 eq), 4-fluorocinnamic acid (1.5 eq).

As shown in Scheme 5, 20-O-trityl- 4α -phorbol and 20-O-trityl- 4α -4-deoxyphorbol were synthesized following a procedure similar to that for 20-O-trityl-phorbol. Compounds 10 (75%, colorless oil) and 11 (80%, colorless oil) were separately synthesized by esterification of 20-O-trityl- 4α -phorbol and 20-O-trityl- 4α -4-deoxyphorbol with decanoic acid (3.0 eq), following a similar method to the synthesis of compound 1b.

Synthesis of target compounds 4c, 6a, 6b, 9a, and 9b

As shown in Scheme 6, 1.2 mmol 6-bromohexanoic acid, 1.5 mmol EDCI and 1.0 mmol DMAP were added into the DCM (12 mL) solution of phorbol-13,20-diacetate (3, 0.42 mmol) and stirred overnight; and then the crude product (0.42 mmol), uracil (3 eq), and K_2CO_3 (1 eq) were dissolved in N,Ndimethylformamide (DMF) and stirred at 60 °C for 12 h to produce intermediate A and the compound 9a (32%) as white amorphous powders. 1.2 mmol 6-bromohexanoic acid or 9bromononanoic acid, 1.5 mmol EDCI, and 1.0 mmol DMAP were added into the DCM (12 mL) solution of 20-trityl-phorbol (1, 0.42 mmol), then stirred overnight and further esterified with acetic anhydride and hydrolyzed C20 protection group to produce crude product. This crude product (0.42 mmol), uracil (3 eq) or piperazine (4 eq), and K₂CO₃ (1 eq) were dissolved in DMF and stirred at 60 or 50 °C for 12 h to produce intermediate B₁-B₃. Oleanolic acid or glycyrrhetic acid (5 mmol), 1,4-dibromobutane (6 eq) and K₂CO₃ (1 eq) were dissolved in DMF and stirred at 60 °C for 12 h to yield intermediate C. As shown in Scheme 7, intermediate A or $\mathbf{B_1}$ - $\mathbf{B_3}$ (0.5 mmol), intermediate C (3 equiv), K_2CO_3 (1 eq), and KI (0.2 eq) were dissolved in methyl cyanide or DMF and stirred at 75 °C for 36 h to yield the compounds 4c (11%), **6a** (35%), **6b** (34%) and **9b** (21%) as white amorphous powders, respectively.

Data of target compounds

Compound **1a**, colorless oil; HR-MS (ESI): m/z, Calcd. for $C_{38}H_{60}NaO_{8}$ [M + Na]⁺: 667.4186, Found 667.4196. ¹H NMR (600 MHz, CDCl₃) δ : 7.55 (1 H, s, H-1), 5.70 (1H, s, -OH), 5.66 (1 H, s, H-7), 5.37 (1 H, d, J = 10.3 Hz, H-12), 4.01 (1 H, d, J = 12.8 Hz, H-20), 3.94 (1 H, d, J = 12.8 Hz, H-20), 3.24 (1 H, s, H-8), 3.23 (1 H, s, H-10), 2.59 (1 H, d, J = 18.9 Hz, H-5 β), 2.47 (1 H, d, J = 19.0 Hz, H-5 α), 2.33–2.25 (4 H, m, -COCH₂- × 2), 2.15–2.12 (1 H, m, H-11), 1.74 (3 H, s, H-19), 1.61–1.57 (4 H, m, -COCH₂CH₂- × 2), 1.26–1.23 (23 H, m, -CH₂- × 10, -CH₃), 1.18 (3 H, s, H-17), 1.04 (1 H, d, J = 4.7 Hz, H-14), 0.85 (9 H, brs, -CH₃ × 3). ¹³C NMR (150

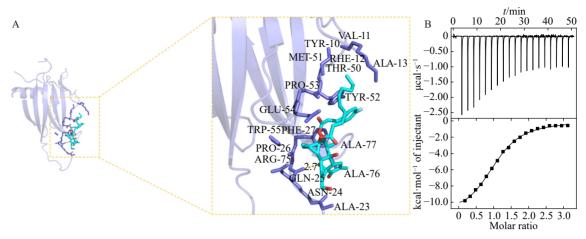


Fig. 5 Effects of compound 3c on PKC. (A) Molecular docking; (B) PKC- δ protein affinity constant (K_d) of compound 3c.

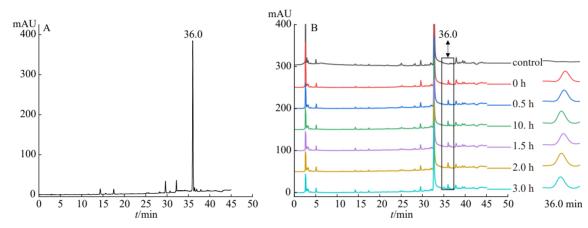


Fig. 6 Stability of compound 3c in plasma in vitro. (A) HPLC of compound 3c; (B) HPLC of compound 3c in rat plasma at different reaction time points.

MHz, CDCl₃) δ : 209.40, 176.48, 173.70, 160.98, 140.77, 132.89, 129.29, 78.48, 76.68, 73.76, 68.08, 65.51, 56.12, 42.98, 39.05, 38.50, 36.42, 34.67, 34.50, 31.91, 31.88, 29.34, 29.28, 29.26, 29.18 (2 × C), 29.09, 25.81, 25.32, 24.62, 23.94, 22.72 (2 × C), 16.94, 14.52, 14.17, 14.16, 10.17.

Compound 2a, a white amorphous powder; HR-MS (ESI): m/z, Calcd. for $C_{28}H_{39}BrNaO_8 [M + Na]^+$: 605.1726, found 605.1713. ¹H NMR (600 MHz, CDCl₃) δ: 7.57 (1 H, s, H-1), 5.66 (1 H, d, J = 4.8 Hz, H-7), 5.65 (1 H, s, -OH), 5.35 (1 H, d, J = 10.3 Hz, H-12), 4.02 (1 H, d, J = 12.9 Hz, H-20),3.96 (1 H, d, J = 12.9 Hz, H-20), 3.40 (2 H, t, J = 6.8 Hz,BrCH₂-), 3.24 (2 H, m, H-8, H-10), 2.56 (1 H, d, J = 19.0 Hz, H-5 β), 2.48 (1 H, d, J = 19.0 Hz, H-5 α), 2.39–2.31 (2 H, m, -COCH₂-), 2.16–2.13 (1 H, m, H-11), 2.06 (3 H, s, -COCH₃), 1.99-1.95 (2 H, m, BrCH₂CH₂-), 1.74 (3 H, d, J = 1.5 Hz, H-19), 1.68-1.63 (2 H, m, -COCH₂CH₂-), 1.50-1.45 (2 H, m, -CH₂), 1.23 (3 H, s, H-16), 1.21 (3 H, s, H-17), 1.06 (1 H, d, J = 5.2 Hz, H-14), 0.87 (3 H, d, J = 6.5 Hz, H-18). ¹³C NMR (150 MHz, CDCl₃) δ : 209.27, 176.22, 171.07, 160.86, 140.76, 133.02, 129.24, 78.41, 77.11, 73.79, 68.09, 65.68, 56.19, 43.07, 39.11, 38.61, 36.50, 34.20, 33.61, 32.49, 27.67, 26.00, 24.02, 23.79, 21.16, 16.87, 14.56, 10.24.

Compound 3c, a white amorphous powder; HR-MS

(ESI): m/z, Calcd. for $C_{39}H_{51}FNaO_8$ [M + Na]⁺: 689.3466, found 689.3474. ¹H NMR (600 MHz, CDCl₃) δ : 7.62 (1 H, d, J = 16.2 Hz, -CH=CH-), 7.60 (1 H, s, H-1), 7.51 (2 H, dd, J =8.6, 5.3 Hz, -Ph) 7.08 (2 H, t, J = 8.5 Hz, -Ph), 6.35 (1 H, d, J = 16.0 Hz, -CH=CH-), 5.76 (1 H, s, -OH), 5.71 (1 H, d, J =4.9 Hz, H-7), 5.51 (1 H, d, J = 10.3 Hz, H-12), 4.05 (1 H, d, J = 10.3 Hz, H-12)J = 12.8 Hz, H-20, 3.99 (1 H, d, J = 12.8 Hz, H-20), 3.29 (1 Hz, H-20)H, t, J = 5.4 Hz, H-8), 3.27 (1 H, s, H-10), 2.59 (1 H, d, J =19.0 Hz, H-5 β), 2.50 (1 H, d, J = 19.0 Hz, H-5 α), 2.42–2.29 (2 H, m, COCH₂-), 2.26-2.21 (1 H, m, H-11), 1.77-1.76 (3 H, m, H-19), 1.65-1.61 (2 H, m, -CH₂-), 1.31-1.26 (15 H, m, $-CH_{2}$ × 6, H-16), 1.22 (3 H, s, H-17), 1.08 (1 H, d, J = 5.2 Hz, H-14), 0.92 (3 H, d, J = 6.5 Hz, H-18), 0.87 (3 H, t, J =6.9 Hz, -CH₃). ¹³C NMR (150 MHz, CDCl₃) δ : 209.22, 176.67, 166.76, 164.08, 160.96, 143.93, 140.65, 133.02, 130.69, 130.12, 129.38, 117.82, 116.22, 78.42, 77.16, 73.83, 68.17, 65.52, 56.28, 43.35, 39.22, 38.72, 36.66, 34.53, 32.00, 29.54, 29.42, 29.39, 29.23, 26.08, 24.69, 24.00, 22.81, 17.07, 14.63, 14.26, 10.27.

Compound 5a, a white amorphous powder; HR-MS (ESI): m/z, Calcd. for $C_{30}H_{43}N_2O_8S$ [M + H]⁺: 591.2740, found 591.2827. ¹H NMR (600 MHz, CD₃OD) δ: 7.62 (1 H, s, H-1), 5.63 (1 H, d, J = 4.8 Hz, H-7), 5.32 (1 H, d, J = 10.1



Hz, H-12), 4.56–4.45 (1 H, m, -N-CH-), 4.32 (1 H, dd, J = 7.6, 4.5 Hz, -N-CH-), 3.97 (2 H, s, H-20), 3.22 (1 H, dt, J = 9.6, 5.0 Hz, -S-CH-), 3.16 (1 H, t, J = 5.4 Hz, H-8), 3.14 (1 H, s, H-10), 2.94 (1 H, dd, J = 12.8, 5.0 Hz, -S-CH₂-), 2.73 (1 H, d, J = 12.8 Hz, -S-CH₂-), 2.56 (1 H, d, J = 19.2 Hz, H-5 β), 2.47 (1 H, d, J = 18.8 Hz, H-5 α), 2.41 (2 H, t, J = 7.3 Hz, COCH₂-), 2.20–2.10 (1 H, m, H-11), 1.77 (3 H, s, H-19), 1.76–1.68 (2 H, m, -CH₂-), 1.63 (2 H, dt, J = 13.5, 7.3 Hz, -CH₂-), 1.50 (2 H, dt, J = 17.5, 8.6 Hz, -CH₂-), 1.22 (3 H, s, H-16), 1.14 (3 H, s, H-17), 0.97 (3 H, d, J = 6.4 Hz, H-18), 0.86 (1 H, d, J = 5.8 Hz, H-14). ¹³C NMR (150 MHz, CD₃OD) δ: 210.56, 176.81, 166.12, 160.85, 142.26, 134.53, 130.64, 86.68, 79.62, 74.73, 68.26, 63.43, 62.15, 61.62, 58.68, 57.05, 44.71, 41.03, 40.03, 38.56, 37.69, 35.24, 29.81, 29.51, 27.86, 26.09, 23.58, 17.86, 15.63, 10.23.

Compound 6a, a white amorphous powder; HR-MS (ESI): m/z, Calcd. for $C_{67}H_{105}N_2O_{11}$ [M + H]⁺: 1113.7718, found 1113.7668. ¹H NMR (600 MHz, CDCl₃) δ: 7.52 (1 H, s, H-1), 5.60–5.58 (2 H, m, H-7, -CH=CH-), 5.27 (1 H, d, J= 3.3 Hz, -OH), 4.09-4.06 (2 H, m, -COOCH₂-), 3.99-3.92 (3 H, m, H-12, H-20), 3.19-3.17 (2 H, m, H-8, -OCH-), 3.10 (1 H, brs, H-10), 1.72 (3 H, s, H-19), 1.33 (3 H, s, H-16), 1.26-1.09 (18 H, -CH₃ × 5, H-17), 1.00-0.96 (6 H, -CH₃, H-18), 0.76 (3 H, s, -CH₃). 13 C NMR (150 MHz, CDCl₃) δ : 209.03, 200.17, 176.79, 176.43, 169.32, 160.44, 140.97, 132.92, 128.79, 128.42, 78.68, 78.35, 77.23, 77.15, 73.45, 67.56, 61.78, 57.89, 56.70, 56.50, 54.90, 52.79 (2 × C), 52.61 $(2 \times C)$, 48.39, 45.38, 44.92, 43.96, 43.19, 41.05, 39.08, 38.90, 38.47, 37.71, 37.05, 35.61, 34.27, 32.73, 31.81, 31.08, 29.08, 28.89, 28.85, 28.55, 28.38, 28.08, 27.34, 27.22, 26.75, 26.45, 26.37, 26.31, 26.26, 24.62 (2 × C), 23.77, 23.37, 23.22, 18.65, 17.46, 16.93, 16.34, 15.59, 15.05, 14.16, 10.14.

Compound 9a, a white amorphous powder; HR-MS (ESI): m/z, Calcd. for $C_{64}H_{84}N_2O_{20} [M + Na]^+$: 1223.5515, found 1223.5513. ¹H NMR (600 MHz, CDCl₃) δ : 7.59 (2 H, s, H-1 \times 2), 7.07 (1 H, d, J = 7.8 Hz, -CH=CH-), 5.70–5.68 (3 H, m, H-7 \times 2, -CH=CH-), 5.52 (2 H, -OH \times 2), 5.38 (2 H, d, J = 10.3 Hz, H-12 × 2), 4.48-4.42 (4 H, m, H-20 × 2), 3.88-3.85 (2 H, m, -NCH₂-), 3.71-3.69 (2 H, m, -NCH₂-), 3.26-3.23 (4 H, m, H-8 × 2, H-10 × 2), 2.55 (2 H, dd, J =19.4, 6.2 Hz, H-5), 2.40 (2 H, dd, J = 19.1, 5.6 Hz, H-5), 2.34-2.32 (4 H, m, COCH₂- × 2), 2.13-2.10 (2 H, m, H-11 × 2), 2.09 (3 H, s, -COCH₃), 2.08 (3 H, s, -COCH₃), 2.04 (6 H, s, -COCH₃ × 2), 1.77–1.76 (6 H, m, H-19 × 2), 1.70–1.65 (6 H, m, $-CH_2 - \times 3$), 1.62–1.58 (2 H, m, $-CH_2$ -), 1.42–1.32 (4 H, m, -CH₂- \times 2), 1.22 (3 H, s, H-16), 1.21 (3 H, s, H-16), 1.20 (3 H, s, H-17), 1.20 (3 H, s, H-17), 1.07-1.06 (2 H, m, H-14 \times 2), 0.90 (3 H, d, J = 6.1 Hz, H-18), 0.86 (3 H, d, J = 6.1 Hz, H-18). ¹³C NMR (150 MHz, CDCl₃) δ : 208.76, 208.72, 173.72, 173.63, 173.31, 173.14, 170.76, 170.74, 163.12, 160.61 (2 × C), 151.25, 142.00, 135.67, 135.64, 133.00, 132.94, 132.52, 132.44, 101.58, 78.08, 78.04, 76.87, 76.72, 73.52, 73.48, 69.35, 69.29, 65.49, 65.48, 56.02 ($2 \times C$), 49.58, 42.94, 42.88, 41.08, 39.21, 39.16, 38.79, 38.69, 36.06, 36.01, 34.32, 34.15, 29.66, 28.68, 27.12, 26.30, 25.87, 25.67, $24.70, 24.49, 23.81, 23.77, 21.06 (2 \times C), 20.94 (2 \times C),$ 16.81, 16.75, 14.53, 14.41, 10.12, 10.10.

Compound 10, colorless oil; HR-MS (ESI): m/z, Calcd. for $C_{40}H_{64}NaO_8$ [M + Na]⁺: 695.4515, found 695.4521. ¹H NMR (600 MHz, CDCl₃) δ : 7.06 (1 H, s, H-1), 5.47 (1 H, d, J = 10.3 Hz, H-12), 5.40 (1 H, s, -OH), 5.21 (1 H, s, H-7), 3.92 (2 H, brs, H-20), 3.75 (1 H, d, J = 14.1 Hz, H-5 β), 3.29-3.28 (1 H, m, H-10), 2.37-2.34 (2 H, m, -COCH₂-), 2.33-2.25 (3 H, m, H-5 α , -COCH₂-), 2.17 (1 H, brs, -CH₂-), 1.97 (1 H, brs, -CH₂-), 1.89 (1 H, s, H-8), 1.75 (3 H, s, H-19), 1.70-1.64 (3 H, m, H-11, -CH₂-), 1.62-1.55 (2 H, m, -CH₂-), 1.37-1.25 (22 H, m, -CH₂- × 11), 1.23 (3 H, s, H-16), 1.18 (3 H, s, H-17), 1.06 (3 H, d, J = 6.4 Hz, H-18), 0.88 (6 H, m, -CH₃ × 2). ¹³C NMR (150 MHz, CDCl₃) δ : 210.43, 176.28, 173.42, 155.83, 140.62, 131.88, 126.07, 77.93, 76.71, 75.62, 68.62, 65.14, 55.84, 43.06, 40.94, 36.24, 34.68, 34.51, 34.23, 31.99, 31.96, 29.61, 29.48, 29.43, 29.39, 29.37, 29.33, 29.18, 29.14, 25.42, 25.35, 24.61, 24.21, 22.78 (2 × C), 16.61, 14.21 $(2 \times C)$, 12.00, 10.69.

Compound 11, colorless oil; HR-MS (ESI): m/z, Calcd. for $C_{40}H_{64}NaO_7$ [M + Na]⁺: 679.4550, found 679.4577. ¹H NMR (600 MHz, CDCl₃) δ : 7.00 (1 H, s, H-1), 5.41 (1 H, d, J = 10.3 Hz, H-12), 5.17 (1 H, s, -OH), 5.07 (1 H, brs, H-7), 3.95 (1 H, d, J = 12.5 Hz, H-20), 3.84 (1 H, d, J = 12.5 Hz, H-20)20), 3.45 (1 H, d, J = 6.2 Hz, H-10), 3.39 (1 H, d, J = 15.5Hz, H-5 β), 2.73 (1 H, t, J = 7.0 Hz, H-4), 2.41 (1 H, dd, J =15.5, 4.8 Hz, H-5 α), 2.35–2.18 (5 H, m, -COCH₂- × 2, H-8), 1.91 (1 H, brs, H-11), 1.73 (3 H, s, H-19), 1.65-1.50 (6 H, m, $-CH_2-\times 3$), 1.37–1.16 (22 H, m, $-CH_2-\times 11$), 1.15 (3 H, s, H-16), 1.12 (3 H, s, H-17), 1.03 (3 H, d, J = 6.3 Hz, H-18), 0.82 $(6 \text{ H, m, -CH}_3 \times 2), 0.73 (1 \text{ H, d, } J = 4.9 \text{ Hz, H-14}).$ ¹³C NMR (150 MHz, CDCl₃) δ : 213.09, 176.05, 173.33, 156.11, 143.25, 136.92, 126.15, 78.00, 75.31, 69.12, 64.89, 49.52, 47.29, 43.08, 40.62, 36.99, 34.51, 34.35, 31.81, 31.78, 29.43, 29.29, 29.25, 29.21, 29.15, 29.00, 28.94, 25.19 (2 × C), 25.15, 25.05, 24.42, 24.08, 22.60 (2 × C), 16.33, 14.04 (2 × C), 11.82, 10.37.

Cytotoxicity Assay

C8166 cell suspension (100 μ L, 4 \times 10⁵/mL) was mixed with different drug solutions, establishing triplicate wells for each treatment. Control wells without any drug treatment were also set up, with AZT (Lot: D1115A, MeilunBio, Dalian, China) serving as the positive control drug. The cultures were incubated at 37 °C in a 5% CO₂ environment for 3 d. Post-incubation, cytotoxicity was detected using the MTT assay. The absorbances at 570 and 690 nm were recorded using an ELx800 microplate reader (Bio-Tek Instruments, Winoski, VT, USA). The 50% cytotoxic concentration (CC₅₀) was subsequently calculated ^[4].

Anti-HIV-1 activity assay

C8166 cells (8 × 10 5 /mL) were infected with HIV-1 $_{\rm IIIB}$ using a multiplicity of infection (M.O.I) of 0.01. After 4-h infection, cells were washed twice with PBS and then resuspended in fresh medium (4 × 10 5 /mL). This suspension was added to 96-well plates with gradient diluted compounds. 3TC (Lot: M20220A, MeilunBio, Dalian, China) was the positive control drug. The plates were incubated at 37 °C in a humidi-

fied incubator with 5% CO_2 for 72 h. Post-incubation, cell culture supernatants were harvested for the subsequent enzyme linked immunosorbent assay (ELISA)^[4].

ELISA for HIV-1 p24 antigen

HIV-1 p24 antigen in cell-free culture medium was measured using an antigen capture ELISA assay. Briefly, antip24 McAb P6F4 (homemade by our laboratory) was added to 96 well microtiter plates coated with Fc-specific anti-mouse IgG (Merck KGaA, Darmstadt, Germany). Triton X-100 (Merck KGaA, Darmstadt, Germany)-treated cell culture supernatant was then added to the wells. The plates underwent sequential incubations: first with diluted rabbit anti-p24 serum, followed by HRP-labeled goat anti-rabbit IgG (Merck KGaA, Darmstadt, Germany). Finally, the OPD substrate solution was added. The optical density of the plates was read at 490/630 nm using an ELISA reader after terminating the colorimetric reaction by H₂SO₄. The percent inhibition of p24 antigen expression was calculated, and the concentration resulting in a 50% reduction in p24 antigen expression (EC₅₀) was calculated [18, 19].

Inhibition of syncytia formation assay

The cytopathic effect (CPE) of HIV-1 on C8166 cells was measured by the Johnson & Byington method. The Tissue Culture Infectious Dose 50 (TCID 50) was determined according to the Reed & Muench method. A suspension of C8166 cells (4 \times $10^5/\text{mL})$ was mixed with HIV-1 $_{\rm IIIB}$ (2000 TCID 50/well) and treated with different concentrations of the compounds (100 μL each). The positive control drugs used were 3TC and T-20 (Lot: M20220A, Roche Laboratories Inc., Basel, Switzerland). Cultures were incubated at 37 °C in a 5% CO2 environment for 72 h. The number of syncytia was counted under an inverted microscope, and the concentration causing a 50% reduction in syncytia formation (EC50) was calculated $^{[19,20]}$.

Detection of HIV-1 replication intermediate products

C8166 cells were treated with either compound 3c or EFV (Lot: A1212AS, MeilunBio, Dalian, China), followed by infection with HIV-1_{IIIB} for 24 h, Subsequent to the infection, the cells were harvested for total DNA extraction. HIV-1 reverse transcriptional intermediate products (ssDNA and 2LTR) were detected by quantitative PCR on an ABI PRISM 7500 Fast Real-time PCR system (Applied Biosystems, Waltham, MA, USA) using primers and probes of ssDNA (Forward primer: 5'-GCCTCAATAAAGCTTGCCTTGA-3', reverse primer: 5'-TGACTAAAAGGGTCTGAGGGATCT-3', probe: 5'-FAM-AGAGTCACACAACAGACGGGCACA-CACTA-TAMRA-3') and 2LTR (Forward primer: 5'-GCCT-GGGAGCTCTCTGGCTAA-3', reverse primer: 5'-AGGTA-GCCTTGTGTGTGGTAGATCC-3', probe: 5'-FAM-TAGT-GTGTGCCCGTCTGTTGTGTGAC-TAMRA-3'). The PCR protocol involved 40 cycles, consisting of pre-denaturation at 95 °C for 2 min, denaturation at 95 °C for 30 s, and and annealing-extension at 60 °C for 30 s [20].

Statistical analysis

The experimental data were visualized using GraphPad

Prism 9.0.0 (GraphPad Software, Boston, MA, USA). The EC_{50} and CC_{50} were calculated by the Reed & Muench method. The therapeutic index of the anti-HIV-1 activity was calculated using the formula $TI = CC_{50}/EC_{50}$.

Molecular docking analyses of compound 3c and PKC-δ

Protein Data Bank (PDB, Research Collaboratory for Structural Bioinformatics, https://www.pdbus.org/) was used to obtain the crystal structure of PKC-δ. The 3D structure of compound 3c was optimized using ChemBio3D Ultra 16.0.0.82 (PerkinElmer, Waltham, USA). This optimization included hydrogenation, charge calculation, and distribution, performed with The Autodock Tool 1.5.6 (ADT, Scripps Research, La Jolla, USA). The molecular docking and estimation of the binding affinities of ligands with proteins were performed using the AutoDock Vina 1.1.2 (The Center for Computational Structural Biology, La Jolla, USA). The final protein-ligand complexes, including cartoon structures and 2D interactions, were visualized using PyMol 2.5.2.0 (DeLano Scientific LLC, San Carlos, USA) [21].

ITC of human-PKC- δ protein binding constants of compound 3c

The PKC-δ protein, expressed and purified *as per* established protocols ^[22], was assessed using an iTC-200 microcalorimeter (MicroCal, Inc, USA) at 25 °C. For the ITC measurements, the protein solution was prepared in 20 mmol·L⁻¹ Tris-Cl (pH 7.5) with 50 mmol·L⁻¹ NaCl (ITC buffer), and the concentration of DMSO was adjusted to 0.5%. Compound 3c, initially prepared in DMSO, was diluted in the ITC buffer to achieve the desired concentrations. The experiment utilized 50 mmol·L⁻¹ protein and 1.0 mmol·L⁻¹ compound 3c. The protein was placed in the cell chamber, and compound 3c, loaded in the syringe, was injected in 20 successive intervals of 150 s each. The obtained data were fitted using the single-site binding model within the origin software package (MicroCal, Inc, USA).

Stability of compound 3c in rat plasma in vitro

Blood samples were obtained from the femoral artery of male rats, anticoagulated with heparin sodium, and centrifuged at 4 °C, 3000 × g for 10 min), and the upper plasma was obtained. The stock solution of P63 was prepared at 1 mg·mL⁻¹ in methanol. To assess stability, we mixed 198 μL of blank plasma with 2 μL of 1 mg·mL⁻¹ compound 3c methanol solution, and the mixture was then incubated for at 37 °C 0, 0.5, 1.0, 1.5, 2.0, 3.0 h, respectively. 400 μL of cold-ice methanol was added to quench the reactions. Then the samples were vortex-mixed and centrifuged (4 °C, 10 $000 \times g$, 10 min). The supernatant was taken, filtered and transferred to vials for HPLC analysis. The blank plasma control group was treated similarly except that 2 µL of methanol instead of compound 3c was used. HPLC was performed using an Agilent 1260 HPLC system (Agilent Technologies, USA) and a JADE-PAK ODS-AQ column (250 mm × 4.6 mm I.D., 5 µm) (Echway, China) at room temperature. The mobile phase was a mixture of methanol (A) and water (B) with gradient elution (0 min: 30% A, 30 min: 90% A, 45 min: 100% A). The

flow rate was 1 mL min⁻¹, the detection wavelength was set at 234 nm, and the injection volume was 10 μ L. Each sample underwent triplicate analyses ^[23].

Conclusions

In conclusion, a series of phorbol esters were innovatively designed and synthesized, leveraging the concepts of privileged substructure and pharmacophore merging. These compounds were evaluated for their anti-HIV-1 activities. The findings revealed that the anti-HIV-1 efficacy of phorbol esters varied depending on the nature and position of the esterification groups. Notably, 13-decanoyl-phorbol-12-transcinnamoyl derivatives and 12,13-didecanoyl-phorbol demonstrated superior anti-HIV-1 activities compared to the control compound, AZT. Particularly significant was the enhanced anti-HIV-1 activity observed in 12-trans-cinnamoyl phorbol derivatives substituted with para electron withdrawing group (F, Cl, or Br). These derivatives also exhibited remarkable inhibition activities on syncytia formation with a high safety index. Thus, this study provides evidence that 13-decanoylphorbol-12-trans-cinnamoyl derivatives with the para electron withdrawing groups in the cinnamoyl moiety and 12,13didecanoyl-phorbol, especially the most active compound 3c, are promising candidates for further preclinical trials. Compound 3c not only inhibits HIV-1 replication and itst entry into cells but also shows potential as a natural activator of PKC.

Supporting Information

Supporting information of this paper can be requested by sending E-mails to the corresponding authors.

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