

•Review•

Natural products: potential therapeutic agents for atherosclerosis

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[ABSTRACT] Atherosclerosis (AS) is an invisible killer among cardiovascular diseases (CVD), which has seriously threatened the life of quality. The complex pathogenesis of AS involves multiple interrelated events and cell types, such as macrophages, endothelial cells, vascular smooth muscle cells and immune cells. Currently, the efficacy of recommended statin treatment is not satisfactory. Natural products (NPs) have attracted increasing attention with regard to their broad structural diversity and biodiversity, which makes them a promising library in the demand for lead compounds with cardiovascular protective bio-activity. NPs can preclude the development of AS by regulating lipid metabolism, ameliorating inflammation, stabilizing plaques, and remodeling the gut microbiota, which lays a foundation for the application of NPs in clinical therapeutics.

[KEY WORDS] Atherosclerosis; Cardiovascular diseases; Pathogenesis; Target; Natural products

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Introduction

Cardiovascular diseases (CVD) is one of the biggest contributors to the burden of chronic diseases, and has replaced infectious diseases as the leading killer around the world [1]. Data from the *Report on Cardiovascular Health and Disease in China 2019* indicated that 330 million Chinese people suffered from CVD with a continuously rising prevalence. Moreover, CVD remained the leading cause of death in 2017, surpassing oncology and other diseases. Atherosclerosis (AS), a chronic inflammatory, immune [2] and epigenetic [3] disease manifested by excessive deposition of low-density lipoprotein (LDL), is a pivotal culprit of ischemic stroke, myocardial infarction and other life-threatening acute cardiovascular events [4]. The development of AS involves multiple events, including oxidation of LDL (ox-LDL), formation of foam cells, migration of vascular smooth muscle cells, activated immune responses, plaque rupture as well as thrombosis. The progress of early AS often goes unnoticed. However, once a plaque ruptures and enters the bloodstream,

it becomes a time bomb that may threaten life at any time. Although many theoretical studies have been conducted to explain the pathogenesis of AS, such as the classical lipid storage hypothesis [5], the inflammatory hypothesis [6-8] and the emerging bone-vascular axis hypothesis [9], the autoimmune hypothesis [5], and even the human papillomavirus (HPV) infection hypothesis [10], the pathological mechanisms of AS are not completely understood. Currently, statins are recommended as the first-line therapy, supplemented with vasodilators and anticoagulants, both of which however do not exert direct inhibitory effect on the development of atherosclerotic plaques. It should be noted that increasing attention has been drawn on the therapeutic targets of AS [11-14], which lays a foundation for drug exploration based on new mechanisms.

Traditional Chinese medicine is a comprehensive medical system where patients are treated with natural plants, animals and mineral remedies. In recent decades, due to the increased interests in natural products (NPs), more and more natural compounds have been demonstrated to exhibit anti-AS and other potential cardiovascular activities. Nevertheless, elucidation of the mechanisms of action has just started, based on cellular models for screening anti-atherogenic NP [15]. Potential active NPs against AS have been discussed in terms of modulating foam cell formation [16] and lowering ox-LDL [17, 18]. In the current review, we summarized the key events and potential targets in the pathogenesis of AS, and systematically discussed promising drug candidates from dif-

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ferent aspects related to disease progression.

Pathogenesis of Atherogenic and Potential Therapeutic Targets

AS is a chronic inflammation and epigenetic disease mediated by excessive LDL, monocyte-derived macrophages, arterial wall endothelial cells and immune cells [19]. It may eventually result in the formation and deterioration of plaques, which then protrudes toward the ruptured blood vessel, enters into the bloodstream and finally generates a thrombus [20]. Some risk factors, such as obesity, smoking, hypertension, and diabetes, accelerate AS driven by LDL, though the underlying mechanisms remains elusive [21]. In addition, the molecular mechanism and pathogenesis of plaque formation in the later stage of disease development are not fully understood. However, every event involved in disease progression, comprising excessive accumulation and oxidation of LDL, foam cell formation, activation of inflammatory response, and plaque rupture caused by endothelial cell damage, can provide potential targets for the treatment of AS.

Accumulation of ox-LDL

Oxidative modification of LDL is a crucial factor in the etiology of AS. Many enzymatic systems mediate the production of reactive oxygen species (ROS), including nicotinamide adenine dinucleotide phosphate oxidase (NOX), lipooxygenase (LO), myeloperoxidase (MPO), monooxygenase, cyclooxygenase and xanthine oxidase. The NOX system is the main source of ROS in the vessel wall and endothelial cells [22]. Inhibition of NOX activity by S17834, a benzo(b)pyran-4-one derivative, significantly relieved aortic atherosclerotic lesions [23]. Additionally, LO is closely related to the synthesis of ox-LDL and studies have verified that disruption of the 12/15-LO gene diminishes AS in ApoE^{-/-} mice [24, 25]. MPO is an abundant heme protein secreted from activated phagocytes and has been reported to play a pivotal role in LDL oxidation during AS [26]. It is also believed that MPO contributes to endothelial dysfunction, thereby acting as a potential multifunctional target [27].

Cholesterol uptake and efflux

The maintenance of cholesterol homeostasis in macrophages is essential to prevent the formation of foam cells, which constitutes the predominant source of plaque development, so as to stimulate the generation of fatty streaks and atherosclerotic plaques. Both excessive uptake and obstruction of the elimination process are unfavorable [28]. Scavenger receptors (SRs) on the surface of macrophages mediate the influx of cholesterol in the form of ox-LDL. Nitro-oleic acid (NO₂-OA) specifically interacts with CD36, a high-affinity SR, competitively restricts the uptake of ox-LDL and reverses cholesterol deposition in cells [29]. Except for CD36, SR-A (CD204) and lectin-like ox-LDL receptor-1 (LOX-1) are the other two primary SRs. However, SR-A may act as a double-edged sword. Studies have shown that inhibition of ox-LDL uptake by macrophages through the LOX-1-NF-κB (nuclear factor-kappa B) pathway mitigates AS in ApoE^{-/-} mice [30]. Both free high-density lipoprotein (HDL) in plasma

and ATP binding cassette transporter A1 (ABCA1) on the surface of macrophages are vital carriers to facilitate cholesterol reverse transport [31]. The ApoA-I mimetic peptide synthesized by GOU *et al.* has been demonstrated to inhibit AS through promoting the physiological function of HDL [32]. Protein arginine methyltransferase 2 (PRMT2) has been proved to mediate cholesterol efflux through up-regulating ABCA1 expression, thereby inhibiting foam cell formation [33].

Endothelial cell injury

Endothelial injury and dysfunction are considered as the basis and initial steps for AS [4]. Nitric oxide (NO) from endothelial cells exhibit anti-AS effects through dilating the blood vessels, inhibiting platelet adhesion and aggregation and suppressing the proliferation of smooth muscle cells [34]. Therefore, NO synthase disorder will inevitably lead to endothelial cell dysfunction. Endothelial nitric oxide synthase (eNOS) is the most critical subtype enzyme in the process of NO synthesis [35]. It should be noted that the mainstream drug statins also mechanically activate eNOS through phosphatidylinositol-3-kinase (PI3K)/protein kinase B (PKB/Akt) signals [36]. Once endothelial cell apoptosis occurs under the pathological conditions, the function of endothelial cells as a physical barrier to the blood vessel wall will be destroyed [37]. The progression of AS can be restricted by targeting classic apoptosis targets, such as signal transducer and activator of transcription 3 (STAT3) [38-40] and NF-κB [41, 42]. Many other targets in the process of ox-LDL-induced endothelial cell damage have also been elucidated [43-45].

Inflammation and immunity

AS is defined as a chronic inflammatory disease with an autoimmune property [2]. The expression of vascular endothelial cell adhesion molecules, chemokines and cytokines promotes the infiltration of inflammatory cells in the arterial wall [46]. The NOD-like receptor protein 3 (NLRP3) inflammasome, consisting of multiple proteins, is a cell sensor of innate immunity. This complex plays a key role in inflammation activation by secreting IL-1β and IL-18 via the caspase-1 dependent pathway [47, 48]. Inhibiting the activation of NLRP3 inflammasome with MCC950 can reduce the development of atherosclerotic lesion in ApoE^{-/-} mice. Pre-clinical studies emphasized that neutralization of IL-1β and IL-18 brought out notable regression of AS [49]. Moreover, blockade of tumor necrosis factor (TNF) comes out to be a valid approach in inhibiting the expression of various inflammatory factors, showing the potential to treat AS [50]. Ox-LDL deposited in atherosclerotic lesions can be recognized by Toll-like receptor 4 (TLR4) and CD36 receptors, followed by induction of the transcription of inflammatory cytokines such as IL-12 and IL-23. CD4⁺, CD8⁺ and natural killer T (NKT) cells participate in related inflammation and immune processes through a series of signal transduction cascades [51-53]. Administration of immune checkpoint inhibitors encompassing anti-CTLA-4 and anti-PD-1 antibodies is able to exacerbate T cell-mediated plaque inflammation [54]. On the contrary, stimulation of regulatory T cells can alleviate AS, which was achieved by low-dose IL-2 treatment [55].

Plaque stabilization

Plaque rupture is an immediate flashpoint for the formation of blood clots, blockage of blood vessels, and the ultimate occurrence of acute cardiovascular events. Plaque lipids and tissue factors are exposed to blood components, and initiates the coagulation cascade, before platelet adhesion and thrombosis [20]. Autophagy is extensively involved in the maintenance of plaque homeostasis, which is currently considered to be at least partly beneficial [56]. AMPK inhibits mTORC1 and stimulates ULK1 during glucose deficiency, thus inducing autophagy and slowing down the malignant development of plaques [57]. Autophagy is also a double-edged sword as excessive autophagic death of smooth muscle cells

may in turn promote the development of plaques [56]. Additionally, up-regulation of PCSK6 has been probed to elicit extracellular matrix remodeling by enhancing the activity of matrix metalloproteinases (MMPs), eventually leading to plaque instability and rupture [58].

Potential NPs for AS treatment

As the clinical effect of AS is limited by mainstream therapy, we summarized the natural components with potent anti-AS bioactivity. Due to the potential for developing new anti-AS therapies through harnessing natural constituents, we categorize them in four groups based on their mechanisms of action (Table 1). Events associated with AS and related po-

Table 1 Protective effect of individual natural compounds against AS

Biological function toward AS	No.	Compounds	Results	Ref
Regulating lipid metabolism	(1)	Vitamin E	Decreasing LDL oxidation 30%–40% in subjects only for five months	[60]
	(2)	Vitamin C	Vitamin C intake alleviates the degree of experimental AS induced by periodontitis in rats	[67]
	(3)	Quercetin	Quercetin alleviates systemic OS, and reduces plaque area <i>in vivo</i> ; inhibits ox-LDL induced ROS formation <i>in vitro</i>	[68]
	(4)	Resveratrol	Anti-inflammation, anti-oxidation, relieving endothelial dysfunction, and remodeling lipid metabolism	[119]
	(5)	Epigallocatechin gallate	Reduction of plasma lipid peroxides	[71]
	(6)	Fraxetin	Fraxetin induces antioxidant enzymes through Nrf2/ARE activation	[72]
	(7)	Neferine	Inhibition of Cu ²⁺ -mediated LDL oxidation in mouse peritoneal macrophages	[73]
	(8)	Ginkgetin	Ginkgetin attenuates lipid deposition in the aorta of atherosclerotic rats	[74]
	(9)	Allicin	Allicin exhibits cholesterol-lowering property in male ICR mice	[75]
	(10)	Icariin	Icariin-treatment lowers serum LDL-C levels to 24%	[76]
	(11)	Xanthohumol	Reduction of accumulated cholesterol in the aortic arch and up-regulation of HDL-C	[77]
	(12)	Guggulsterone	Reduction of total serum lipid and total serum cholesterol	[78]
	(13)	Phytanic acid	Induction of hypolipidemia	[79]
	(14)	Aegeline 2	Reduction of plasma triglyceride (Tg) levels by 55%, and total cholesterol (TC) by 24%	[80]
	(15)	Tanshinone IIA	Inhibition of the mRNA expression of CD36 SR-A and PPAR γ in the aorta	[82]
	(16)	Chrysin	Chrysin increases the mRNA levels of PPAR γ , LXR α , ABCA1 and ABCG1	[84]
	(17)	Asperlin	Asperlin inhibits LPS but not ox-LDL-evoked foam cell formation and promotes cholesterol efflux	[85]
	(18)	Ginsenoside Compound K	Up-regulation of ABCA1 and ABCG1 expression in macrophages	[86]
	(19)	Rutaecarpine	Induction of ABCA1 and SR-BI expression in the liver of ApoE ^{-/-} mice	[87]
	(20)	Alpinetin	Alpinetin elevates the mRNA levels of PPAR γ , LXR α , ABCA1 and ABCG1	[88]
	(21)	Baicalin	Stimulation of the PPAR γ -LXR α -ABCA1/ABCG1 pathway	[89]
	(22)	Lycopene	Stimulation of PPAR γ , LXR α and ABCA1 expression	[90]
	(23)	1,2,3,4,6-Penta-O-galloyl- β -D-glucose	Induction the expression of SR-BI and ABCA1 in J774 and THP-1 macrophages	[91]
	(24)	6-Dihydroparadol	Enhancement of ABCA1 and ABCG1 protein levels	[92]
	(25)	Paeonol	Paeonol up-regulates the protein stability of ABCA1	[93]

Continued

Biological function toward AS	No.	Compounds	Results	Ref
Regulating lipid metabolism	(26)	Arctigenin	Arctigenin treatment enhances the expression of ABCA1, ABCG1 and ApoE	[94]
	(27)	α -Asarone	α -Asarone treatment blocks the induction of SR-B1 and promotes the induction of ABCA1 and ABCG1	[95]
	(28)	Chlorogenic acid	Elevation of the mRNA levels of PPARc, LXR α , ABCA1 and ABCG1	[96]
	(29)	Berberine	Inhibition of LOX-1 expression; maintenance of intestinal flora homeostasis	[97,123, 154-156]
	(30)	Piperine	Piperine up-regulates the expression of ABCA1	[98]
	(31)	Leonurine	Up-regulating the expression of PPAR γ , LXR α , ABCA1 and ABCG1	[99]
Fighting inflammation	(32)	Procyanidin B2	Suppressing the activation of NLRP3; and inhibiting caspase-1 activation and IL-1 β secretion	[102]
	(33)	Apigenin	Apigenin inhibits LPS-induced IL-1 β production by inhibiting caspase-1 activation through disruption of NLRP3	[103]
	(34)	Dihydromyricetin	Inhibition of the NLRP3 inflammasome	[104]
	(35)	Hematein	Restriction of ROS generation and NF- κ B activation	[108]
	(36)	7-Hydroxy frullanoide	Inhibition of LPS-induced cytokine production, and the expression of VCAM1, ICAM1 and E-selectin in TNF- α -stimulated HUVECs	[109]
	(37)	Ginkgolic acid	Reduction of intracellular ROS and ox-LDL-induced NF- κ B	[105]
	(38)	Juglanin	Juglanin inhibits the inflammatory response by suppressing the expression of IL-1 β , MCP-1, and HMGB1	[113]
	(39)	Policosanols	Reduction of inflammation markers such as sCD40L, sP-selectin, and IFN- γ	[114]
	(40)	10-Dehydrogingerdione	Reduction of inflammation markers such as sCD40L, sP-selectin, and IFN- γ	[114]
	(41)	Shikonin	Inhibition of inflammatory activated CD4 ⁺ T cells and proinflammatory macrophages in plaques	[115]
	(42)	Kaempferol	Modulation of pro-inflammatory enzyme activities and relative genes	[116]
	(43)	Morelloflavone	Potent ROS scavengers	[117]
	(44)	Volkensiflavone	Effective ROS scavengers	[117]
	(45)	Diosgenin	Anti-inflammation, mitigating endothelial dysfunction and remodeling lipid metabolism	[118]
Stabilizing plaques	(46)	Cryptotanshinone	Inhibition of the expression of LOX-1 and MMP-9, ROS generation and NF- κ B activation.	[122]
	(47)	Ginsenoside F1	Reduction of LOX-1 and TLR4 expression, and MPO distribution	[124]
	(48)	Ursolic acid	Anti-inflammation and anti-oxidation; restriction of LOX-1	[125-127]
	(49)	Hyperoside	Inhibition of oxLDL-induced LOX-1 expression, ERK activation, and cell proliferation	[128]
	(50)	Celastrol	Celastrol suppresses atherosclerotic plaque in ApoE ^{-/-} mice through inhibiting LOX-1 and OS	[129]
	(51)	Trichosanatine	Inhibition of the activated LOX-1/p38 MAPK pathway; protecting HUVECs against ox-LDL-induced injury	[130]
	(52)	Dihydrotanshinone I	Inhibition of LOX-1 mediated by the NOX4/NF- κ B signaling pathway both <i>in vitro</i> and <i>in vivo</i>	[131]
	(53)	N-methylsecoglaucine	Inhibition of rabbit platelet aggregation and the release of ATP induced by arachidonic acid and collagen	[132]
	(54)	Gypenoside	The PI3K/Akt/Bad signaling pathway is activated to modulate the apoptosis-related protein expression in the aorta	[133]
	(55)	Acacetin	Acacetin activates Sirt1/Sirt3/AMPK signals to protect the vascular endothelium	[134]
	(56)	Myricitrin	Reduction of endothelial cell apoptosis <i>via</i> the STAT3 and PI3K/Akt/eNOS signaling pathways; prevention of OS injury	[39,135]
	(57)	Luteolin	Down-regulation of apoptosis through inhibiting MMP9 and activating the PI3K/Akt signaling pathway	[136]
	(58)	Isorhamnetin	Inhibition of macrophage apoptosis by decreasing ROS levels and lipid deposition	[137]

Continued

Biological function toward AS	No.	Compounds	Results	Ref
Stabilizing plaques	(59)	Vitexin	Vitexin increases the expression of p-AMPK and decreases the expression of p-mTOR, thus activating autophagy	[138]
	(60)	Lupeol	Activation of macrophage autophagy through increasing LC3-II levels and inhibiting p62	[139]
	(61)	Curcumin	Inhibition of apoptosis and induction of autophagy via the Akt/mTOR pathway.	[140]
	(62)	Betulinic acid	Betulinic acid induces eNOS expression, thus preventing endothelial dysfunction	[142]
	(63)	Imperatorin	Anti-lesion role in ox-LDL-stimulated VSMCs by inhibiting the PI3K/Akt/mTOR pathway	[145]
	(64)	Securinine	Inhibiting the proliferation and migration of aortic smooth muscle cells in a dose-dependent manner	[146]
	(65)	Ginsenoside Rg3	Stronger antiproliferative and antimigratory effects due to stronger PPAR activation	[147]
	(66)	Artemisinin	Inhibition of MMP-9 expression and activity by suppressing the PKC delta/ERK/p38 cascade	[148]
	(67)	Castanospermine		
	(68)	Isoliquiritin	Potential anti-angiogenesis agents	[149]
Regulating the gut microbiota	(69)	Radicicol		
	(70)	Ferulic acid	Modulation of the ratio of Firmicutes to Bacteroidetes	[157]
	(71)	Chitin-glucan	Modulation of Lactobacillus and Alistipes, thus ameliorating endothelial and inflammatory dysfunctions	[158]
	(72)	Naringin	Modulation of 7 α -dehydroxylase-producing bacteria	[159]
	(73)	Anthocyanin	Up-regulation of the proportions of Lactobacillus, Akkermansia, Bifidobacterium and Roseburia	[160]
	(74)	Trigonelline		[161]
	(75)	2,3,5,4-Tetrahydroxy-stilbene-2-O- β -D-glucoside		[162]
	(76)	Ginkgolide B	Promising gut microbiota regulators	[163]
	(77)	Inulin		[164]
	(78)	Mannan oligosaccharides		[165]

tential natural products are showed in Fig. 1.

Regulating lipid metabolism

Serving as the most fundamental inducement, oxidative metabolism and transport of LDL-cholesterol in blood vessels determine the fate of AS. Hypolipidemic statins are still the main approach to treating AS. However, it remains undesirable in severe cases. Natural compounds that regulate lipid metabolism mainly focus on anti-oxidation, lowering blood lipids, and inhibiting macrophages-derived foam cell formation (Fig. 2).

Anti-oxidation

According to clinical trials, vitamin E (1) exerted certain effect on the reduction of ox-LDL [59, 60]. Ganini and Mason found that α -tocopherol inhibited the production of lipid-radical and protein-derived free-radical caused by Cu²⁺ [61]. Studies of Moser *et al.* suggested that vitamin C (2) was involved in the antioxidant process of LDL [62-64]. Better performance can be obtained especially when vitamin C was used in combination with vitamin E (1) [65, 66]. An experiment using a rat model of AS caused by periodontitis showed that vitamin C (2) alleviated symptoms by decreasing oxidative stress

(OS) [67]. Quercetin (3) is ubiquitous in our diets and has been proved to protect patients suffering from CVD. Quercetin (3) has been reported to suppress the production of ROS through NOX inhibition in the peritoneal macrophages of high-fat diet (HFD) mice [68]. In a male rat model of oxidation damage, quercetin (3) up-regulated paraoxonase 1 activity, preventing LDL from oxidative modification [69]. Both resveratrol (4) and epigallocatechin gallate (5) are potential anti-oxidants with a similar mechanism of action, which have been verified in clinical use [70, 71]. Some compounds that have not been clinically verified such as fraxetin (6) and neferine (7) also exhibited antioxidant activity in preliminary studies [72, 73].

Lowering blood lipids

Ginkgetin (8) has been found to decrease the serum levels of total cholesterol, triglyceride and LDL-cholesterol in atherosclerotic rats [74]. In an experiment exploring the effect of allicin (9) on hypercholesterolemia in male ICR mice, decreases in cholesterol, triglyceride and glucose were observed, indicating its potential value in lowering cholesterol [75]. Icariin (10), the major bioactive component of *Epimedium brevicornum*, also had similar functions in a rab-

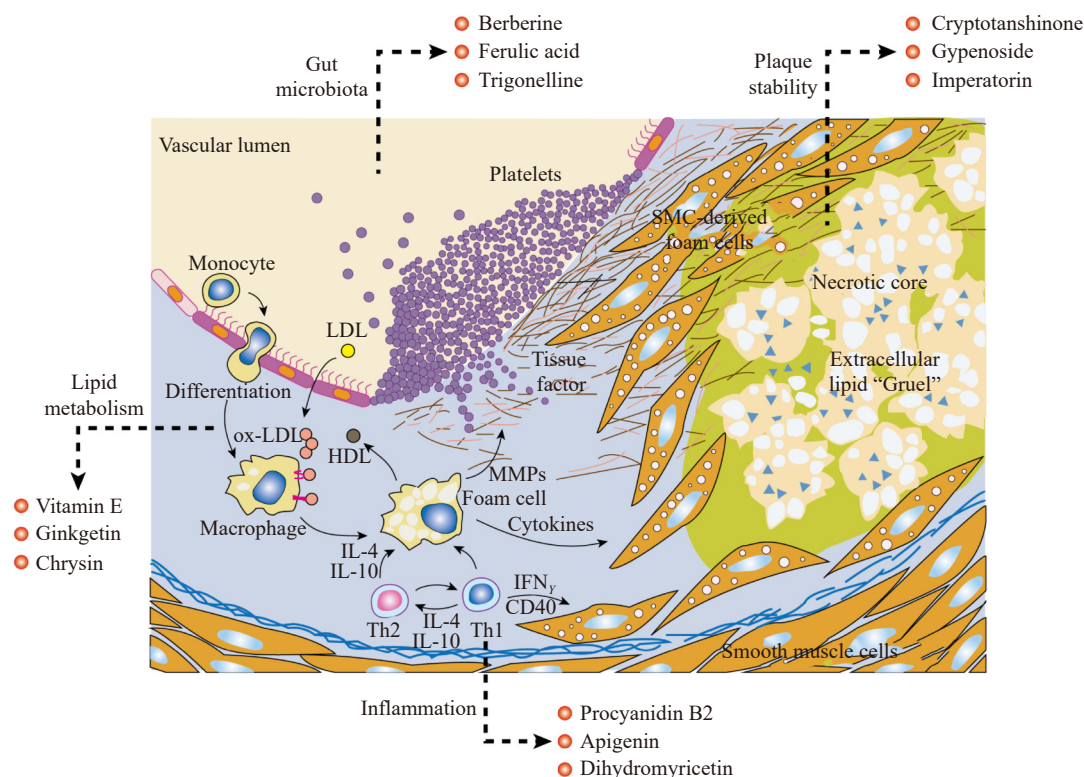


Fig. 1 Events and related potential natural products in AS

bit hyperlipidemia model [76]. Xanthohumol (11) has been reported to suppress cholesterol accumulation in lesion areas by HDL-cholesterol metabolism through ApoE enhancement [77]. In addition to those bioactive ingredients mentioned above, more than a dozen of compounds such as guggulsterone (12), phytanic acid (13) and aegeline 2 (14) have been proved in various cell and animal models to be the candidates for lowering blood lipid [78-81].

Modulating lipid metabolism in macrophages

Cholesterol uptake and excretion by macrophages, which is termed foam cell formation, is a pivotal target of drug intervention. SRs located on the surface of macrophages are responsible for the endocytosis of ox-LDL. TANG *et al.* demonstrated that tanshinone IIA (15) down-regulated the expression of CD36 in ApoE^{-/-} mice, thus mitigating AS [82]. Interestingly, vitamin E (1) was used as a CD36 inhibitor [83]. Chrysin (16), asperlin (17), ginsenoside compound K (18) and rutaecarpine (19) treatments mechanically elevated cholesterol efflux mediated by enhanced ACBA-1 activity [84-87]. Alpinetin (20), baicalin (21) and other alkaloids have been reported to promote cholesterol efflux via the PPARα-LXRα-ABCA1 pathway [88, 89]. Lycopene (22) as a typical compound of carotenoids, suppressed cholesterol synthesis and efflux at the concentration of 10 μmol·L⁻¹ [90]. Phenolic compounds represented by 1,2,3,4,6-penta-*O*-galloyl-β-d-glucose (23), 6-dihydroparadol (24) and paeonol (25) [91-93], phenylpropanoids represented by arctigenin (26), α-asarone (27), chlorogenic acid (28) [94-96] as well as alkaloids represented by berberine (29), piperine (30) and leonurine (31) [97-99] are all

active NP targeting proteins associated with foam cell formation involving ABCA1, ABCG1, HMG-CoA and SR-BI.

Fighting inflammation

Procyanidins is a type of polyphenolic compounds that widely exists in red wine and grape seeds [100, 101]. Procyanidin B2 (32) is the major natural procyanidin, which has been reported to suppress the activation of NLRP3 inflammasome via the activator protein-1 pathway in human umbilical vein endothelial cells (HUVECs), indicating its cardiovascular protective effect [102]. ZHANG *et al.* found that flavonoid apigenin (33) impeded the production of IL-1β and other inflammatory cytokines induced by lipopolysaccharide (LPS) in macrophages. Their further research revealed that the assembly of NLRP3 was destroyed [103]. In addition, dihydromyricetin (34) has been proved to reverse the activation of the NLRP3 inflammasome in HUVECs after palmitic acid treatment, thereby preventing AS through inhibiting pyroptosis [104]. NF-κB is also an important signal molecule in inflammation [105-107]. The effect of hematein (35) in alleviating AS was evaluated using a hyperlipidemia mouse model and results showed that hematein (35) reduced inflammatory mediators by interfering NF-κB activation [108]. 7-Hydroxy frullanoide (36), an active ingredient in *Sphaeranthus indicus*, together with quercetin (3) and ginkgolic acid (37), have been reported to display anti-inflammatory activity through inhibiting the NF-κB pathway [105, 109-112]. Asperlin (17) [85], juglanin (38) [113], policosanol (39) as well as 10-dehydrogingerdione (40) [114] down-regulated the level of inflammatory factors such as TNF-α and IL-1β. Immune cells play an

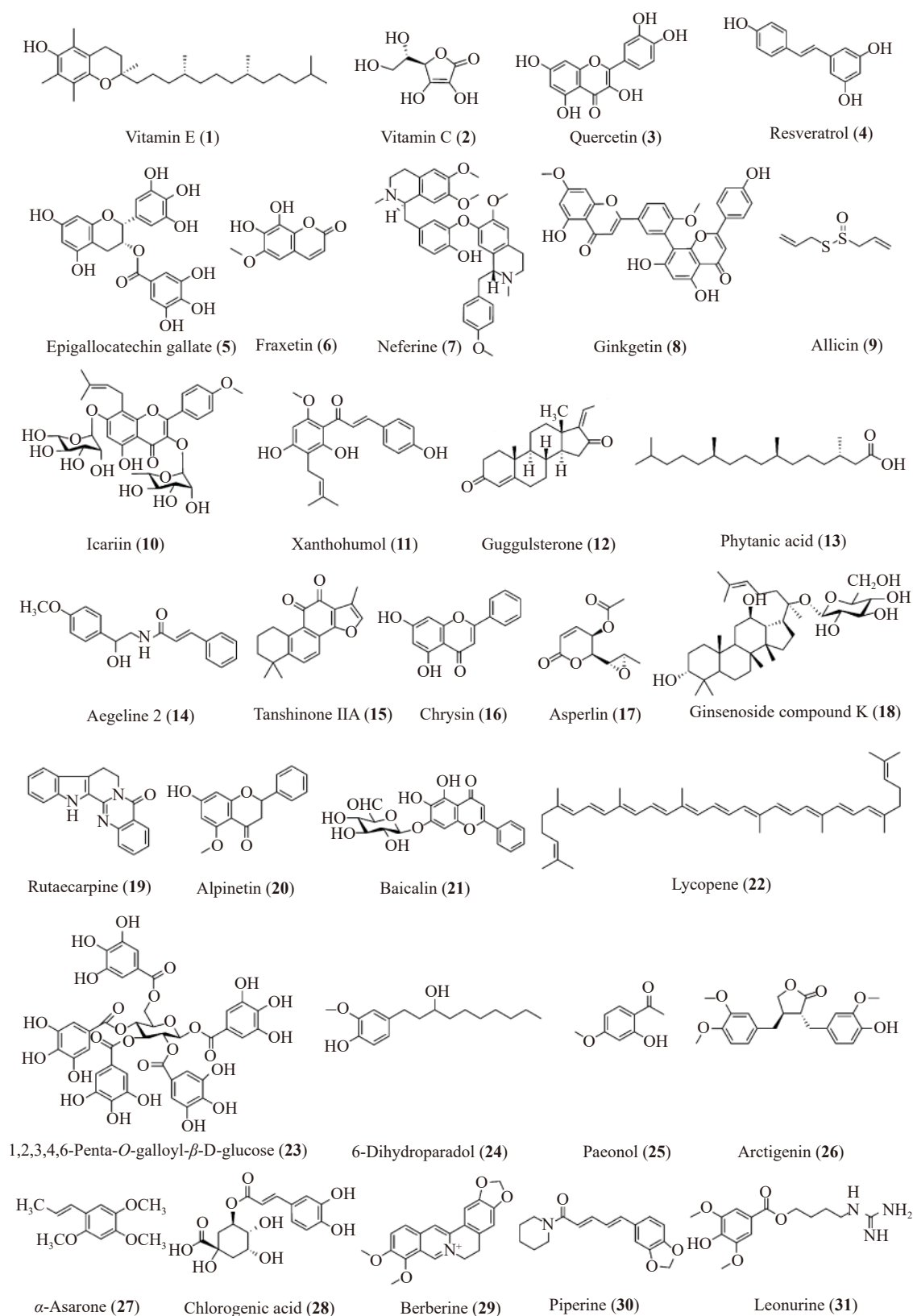


Fig. 2 Natural compounds regulating lipid metabolism against AS

indispensable role in the development of inflammation and are considered to be regulated by shikonin (41)^[115]. Shikonin (41) has been elucidated to repress CD4⁺ T cell inflammatory

activation through inhibiting its essential process of glycolysis and oxidative phosphorylation^[115]. Kaempferol (42) exerted robust anti-inflammation effects due to multiple mechan-

isms, such as anti-oxidation, modulating pro-inflammatory enzyme activity and relative genes, and reducing adhesion molecule expression [116]. Otherwise, morelloflavone (43) and volkensiflavone (44) showed atheroprotective effect as ROS scavengers [117]. Both diosgenin (45) coupled with resveratrol (4), are outstanding natural compounds with huge potential in the treatment of AS, benefiting from multiple functions such as anti-inflammation, relieving endothelial dysfunction, and remodeling lipid metabolism [118, 119]. Natural constituents displaying anti-inflammatory potential are shown in Fig. 3.

Stabilizing plaques

If foam cell accumulation is the cornerstone of lesion formation, plaque rupture is the last trigger for atherosclerotic deterioration leading to myocardial infarction and stroke, though ischemia caused by end-stage luminal stenosis also represents an underlying factor [120]. NPs that maintain plaque stability by precluding platelet adhesion, inducing macrophage and endothelial cell autophagy rather than apoptosis, counteracting angiogenesis and other ways will display broad

cardiovascular benefits in the long term.

Precluding platelet aggregation

LOX-1 is the main receptor in endothelial cells responsible for ox-LDL recognition and internalization [121]. The activation of LOX-1 mediated endothelial dysfunction, leukocyte adhesion, smooth muscle cell proliferation and migration, foam cell formation as well as platelet activation, which constitute the essential events resulting in atherosclerotic plaque rupture [20]. Cryptotanshinone (46) is a dominant diterpenoid with bioactivity extracted from *Salvia miltiorrhiza*. One study in ApoE^{-/-} mice pointed out that cryptotanshinone (46) significantly down-regulated LOX-1 and MMP-9 expression and inhibited ROS generation, thereby maintaining the stability of plaques [122]. Another study in human macrophage-derived foam cells verified that berberine (29) suppressed the elevated expression of LOX-1 caused by ox-LDL in a dose and time-dependent manner [123]. QIN *et al.* investigated that ginsenoside F1 (47) attenuated LOX-1 levels and improved endothelial cell viability damaged by ox-

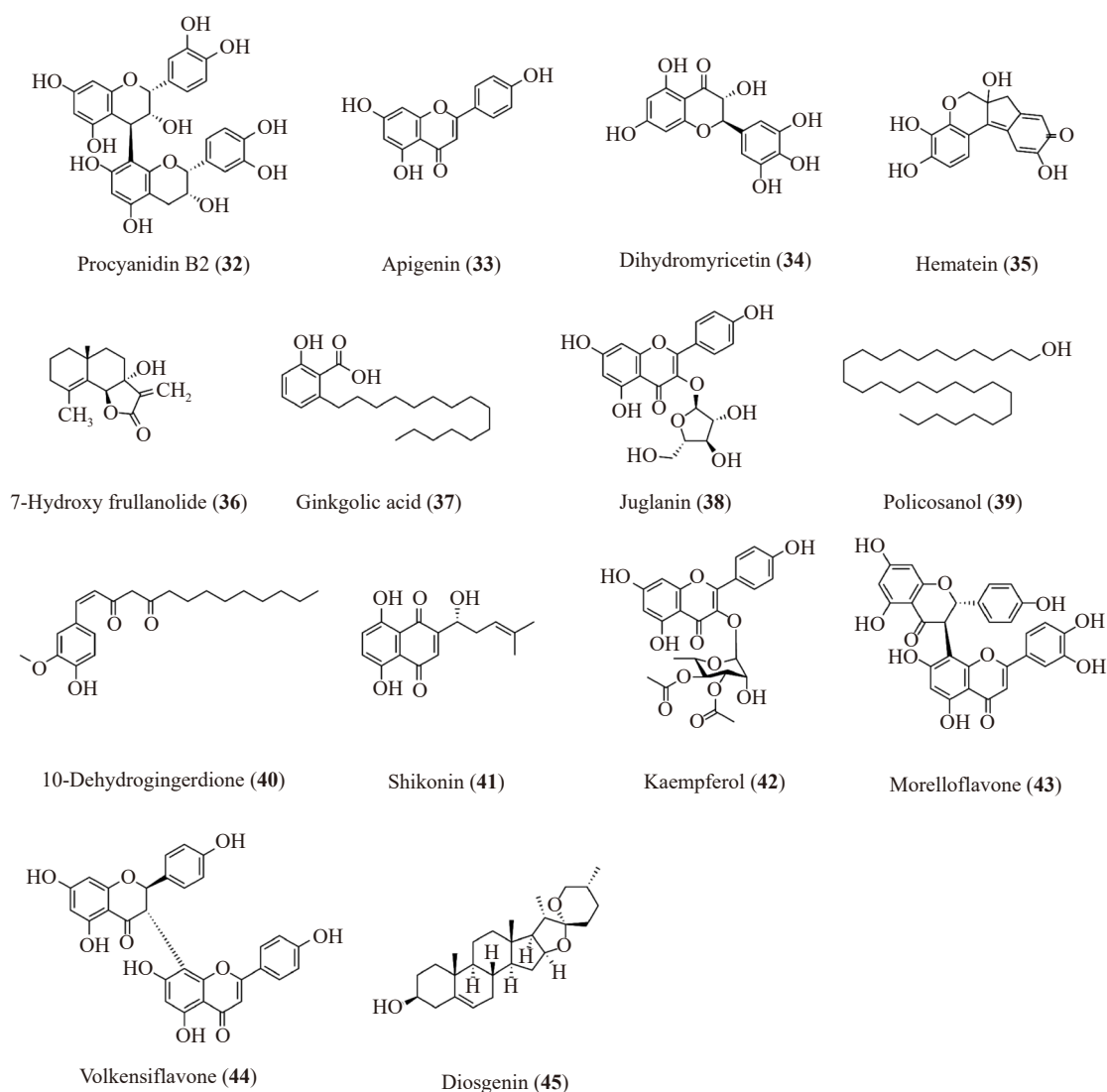


Fig. 3 Natural compounds fighting inflammation in AS

LDL^[124]. Ursolic acid (48) belonging to the class of phytonutrient has been addressed to play a crucial role in anti-inflammation and anti-oxidation^[125, 126]. Ursolic acid (48) (100 mg·kg⁻¹·d⁻¹) markedly lessened necrotic core sizes dominantly through restricting the role of LOX-1^[127]. An additional study utilizing vascular smooth muscle cells showed that hyperoside (49) hampered ox-LDL-induced LOX-1 expression^[128]. Compounds with a similar mechanism of action include celastrol (50), trichosanatine (51) and dihydrotan-shinone I (52)^[129-131]. In addition, five phenanthrene alkaloids represented by *N*-methylsecoglaucine (53) inhibited platelet aggregation induced by arachidonic acid, collagen, or some other stimuli, indicating their cardiovascular protective biologic effects^[132].

Regulating apoptosis/autophagy in endothelial cells and macrophages

Apoptosis of either endothelial cells or macrophages can promote the development of plaques, while autophagy acts the opposite role. Gypenoside (54) has been reported to abolish the apoptosis of endothelial cells by regulating the mitochondria via the PI3K/Akt/Bad axis^[133]. Acacetin (55) was reported to protect HUVECs against apoptosis in an *in vitro* endothelial injury model stimulated by high glucose^[134]. Myricitrin (56) is a natural flavonoid isolated from the root bark of *Myrica cerifera*. Myricitrin (56) treatment has been proved to effectively reduce endothelial cell apoptosis through the STAT3 and PI3K/Akt/eNOS signaling pathways^[39] or prevention of OS injury^[135]. Relevant studies have shown that luteolin (57) and isorhamnetin (58) interfered with macrophage apoptosis, thus reducing plaque instability^[136, 137]. One study in HUVECs designed to address the effect of vitexin (59) on endothelial dysfunction revealed that vitexin (59) induced autophagy, while inhibiting apoptosis^[138]. Lupeol (60) treatment remarkably activated the autophagy of macrophages as indicated by increased LC3-II levels and inhibition of p62^[139]. Curcumin (61) and quercetin (3) were also considered to be autophagy activators of macrophages^[140, 141]. Betulinic acid (62), a natural pentacyclic triterpenoid, has been reported to prevent endothelial dysfunction by promoting eNOS expression through stimulating the HDACS/ERKS/KLF2 pathway^[142].

Inhibiting angiogenesis by targeting MMP-2/9

The proliferation and migration of vascular smooth muscle cells (VSMCs) due to MMP-2 and MMP-9 overexpression are crucial steps in the pathogenesis of intimal thickening and plaque instability^[143, 144]. A recent study has demonstrated that angelica dahurica-derived imperatorin (63) precluded the migration of VSMCs induced by ox-LDL through repressing MMP-2^[145]. Securinine (64) was also identified to attenuate the proliferation and migration of smooth muscle cells by abating the expression of both adhesion molecules and MMP-2/9^[146]. Similar observations were reported after treatment with ginsenoside Rg3 (65), which however worked by activating PPAR γ ^[147]. Artemisinin (66), a classic antimalarial drug, has been explored to de-

crease MMP-9 expression in phorbol myristate acetate-stimulated macrophages, thus exhibiting potential anti-atherosclerotic biologic effects^[148].

With regard to the prevention and treatment of angiogenesis associated with AS, natural angiogenesis inhibitors exhibit markedly high potency^[149]. Castanospermine (67), isoliquiritin (68), radicol (69) and others have been reported to be potentially active compounds. Nevertheless, the specific inhibitory mechanisms of action remain to be explored^[149]. Natural compounds for plaque stability are shown in Fig.4.

Regulating the gut microbiota

The gut microbiota is partially involved in the progression of AS at least by interfering with lipid metabolism and inducing systemic inflammation^[150, 151]. Gut microbiota metabolites such as trimethylamine-*N*-oxide can cause platelet overactivation, dyslipidemia, OS and endothelial dysfunction, which are the key factors to aggravate AS^[152]. Therefore, the gut microbiota becomes an important target in the treatment of AS^[153]. Related active natural compounds are displayed in Fig.5.

Berberine (29) is the main active component of *Rhizoma coptidis*, whose anti-inflammatory and endothelial cell-protective effects in the treatment of AS have been widely investigated^[154]. In HFD-fed ApoE^{-/-} mice, berberine (29) significantly increased the abundance of *Akkermansia* in the gut^[155]. Moreover, further studies showed that berberine (29) significantly changed the composition of the gut microbiota, which varies from different dosages. *Alistipes* and *roseburia* were enriched in the high-dose group while *blautia* and *allobaculum* were enriched in the low-dose group. The microbiota exhibited excellent anti-inflammatory effects, thanks to glucolipid metabolism and the synthesis of short-chain fatty acids^[156]. Ferulic acid (70) is a phenolic acid widely distributed in plants, which has been proved to reverse gut dysbiosis through turning the ratio of Firmicutes/Bacteroidetes to the normal^[157]. Chitin-glucan (71) and pomegranate peel extract were shown to decrease the relative abundance of *Alistipes* and *Lactobacillus*, thus abating endothelial and inflammatory disorders in a mouse CVD model^[158]. As a major flavonoid in Citrus species, naringin (72) has been found to lower cholesterol content in the serum and liver by adjusting the abundance of 7 α -dehydroxylase-producing bacteria^[159]. Anthocyanin (73) significantly up-regulated the proportions of *Lactobacillus*, *Akkermansia*, *Bifidobacterium* and *Roseburia*, thus reversing the “pro-atherogenic” gut microbiota community^[160]. Aside from trigonelline (74), 2,3,5,4'-tetrahydroxy-stilbene-2-*O*- β -D-glucoside (75) and ginkgolide B (76), polysaccharides such as inulin (77) and mannan oligosaccharides (78) are also potential anti-AS ingredients in modulating the gut microbiota^[161-165].

Concluding Remarks and Future Perspectives

AS is the main risk factor of thrombosis, which is the dominating culprit of death worldwide. Despite great progress achieved during the diagnosis and treatment of AS in the past decades, AS is still a global problem threatening hu-

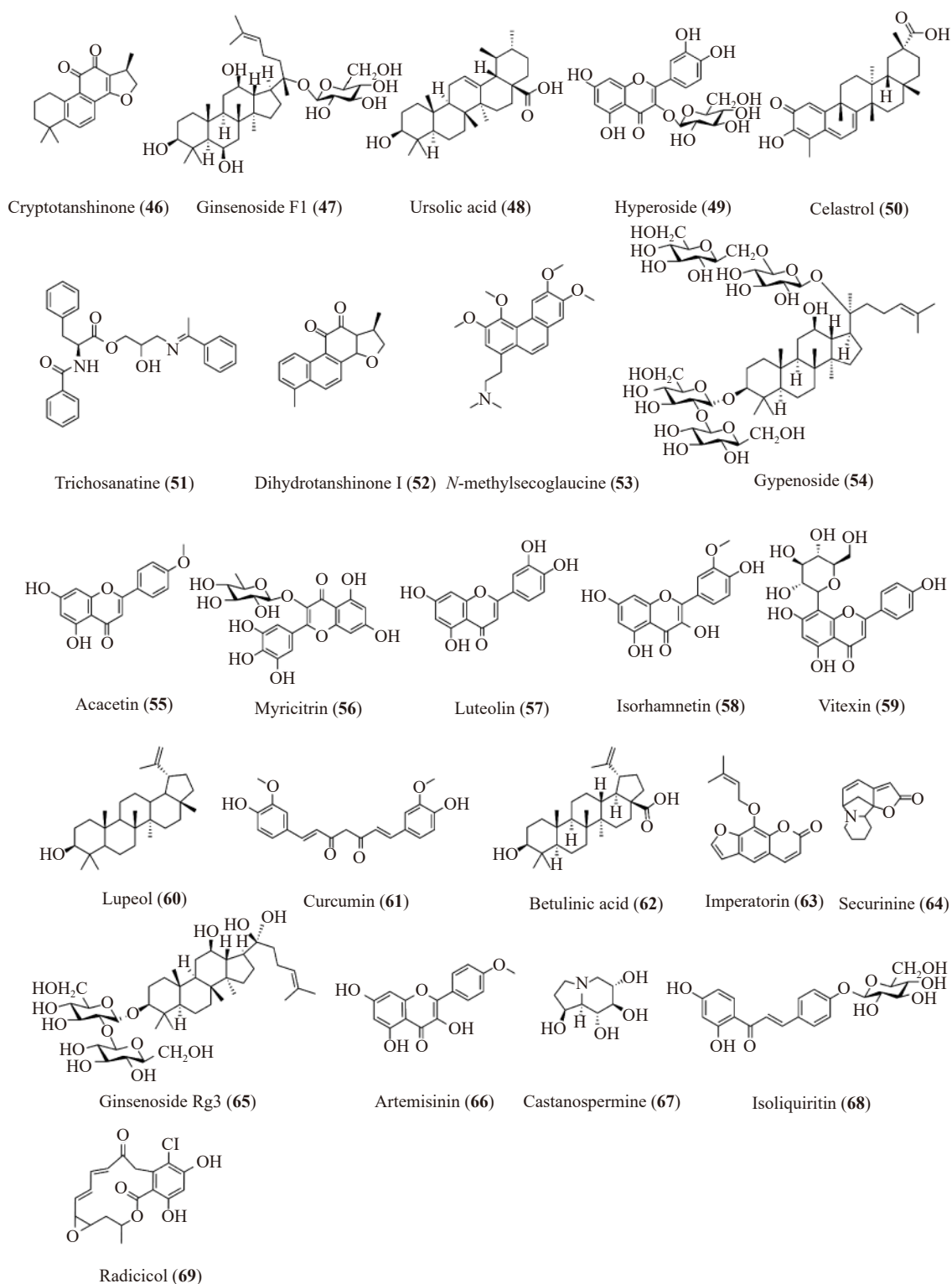


Fig. 4 Natural compounds for atherosclerotic plaque stability

man health. With the deepening of research, the development of a variety of diagnostic and detection technologies has allowed us understand the pathogenesis and treatment targets of AS in a more comprehensive manner. For instance, metabolomics and proteomics are leveraged to investigate the changes of plasma metabolites in animal models of AS, thus helping

to predict biomarkers [166-169]. The application of network pharmacology and bioinformatics has improved our comprehension of the mechanism of action of drugs [170, 171].

NPs with medicinal values are treasures bestowed by nature. The source and structure diversity render them more potent in treating AS and plenty of compounds have been re-

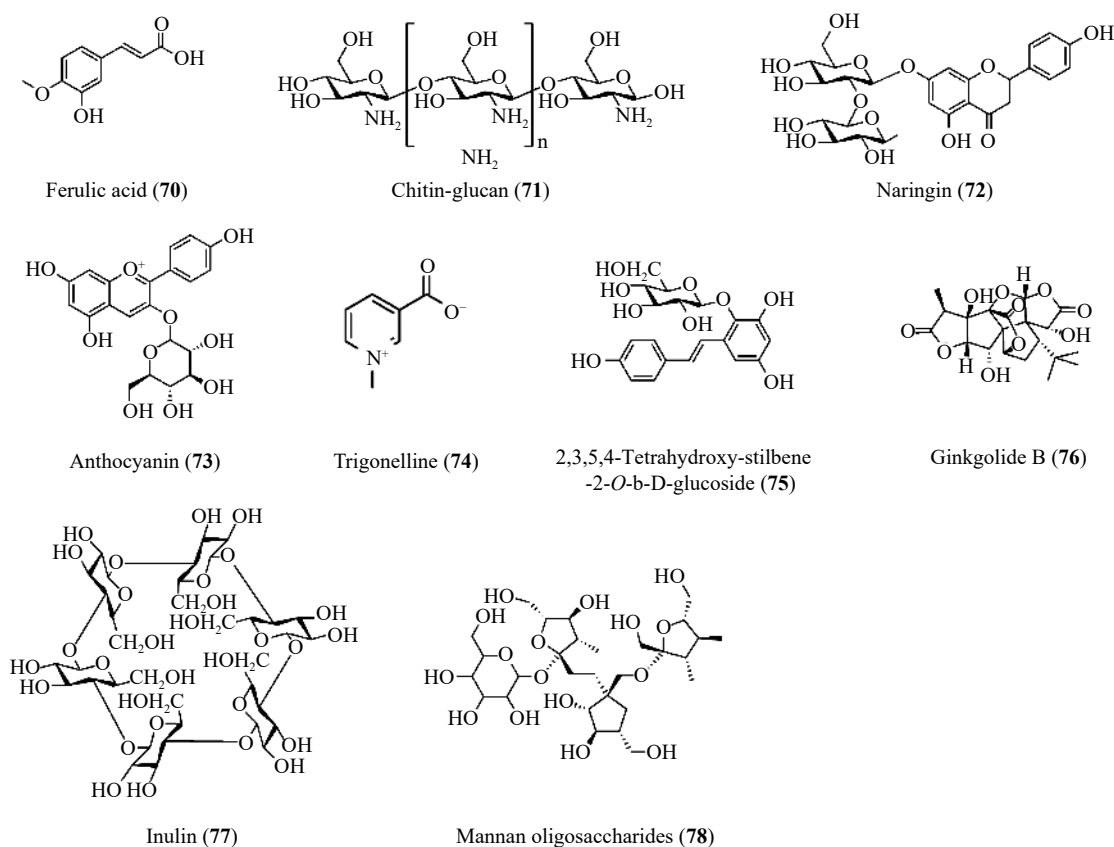


Fig. 5 Natural products for gut microbiota homeostasis

ported to be potent. With regard to the complex and non-independent pathogenesis and development of AS, multi-drug combination based on multiple targets will be of great significance. Such therapy can not only achieve synergistic efficacy but also reduce the untoward effects of certain drugs. Clinical trials have shown that the combination of low-dose rivaroxaban and acetylsalicylic acid to prevent thrombosis is an effective option to treat AS [172]. New technical methods such as microbiology and metabolomics have been utilized to investigate the anti-AS mechanism of natural products and traditional Chinese medicines, encouraging pharmaceutical researchers to energetically screen active ingredients from natural product libraries and explain the target of action of drugs [173]. Moreover, great success has been seen in expanding nanomedicine to the treatment of cardiovascular diseases such as AS [174-176], which also provide a new and powerful approach for the targeted delivery of NPs to the sites of atherosclerotic lesion in the near future.

References

- [1] Yin M, Li C, Jiang J, et al. Cell adhesion molecule-mediated therapeutic strategies in atherosclerosis: from a biological basis and molecular mechanism to drug delivery nanosystems [J]. *Biochem Pharmacol*, 2021, **186**: 114471.
- [2] Kobiyama K, Ley K. Atherosclerosis a chronic inflammatory disease with an autoimmune component [J]. *Circ Res*, 2018, **123**(10): 1118-1120.
- [3] Xu S, Pelisek J, Jin ZG. Atherosclerosis is an epigenetic disease [J]. *Trends Endocrinol Metab*, 2018, **29**(11): 739-742.
- [4] Tabas I, García-Cardeña G, Owens GK. Recent insights into the cellular biology of atherosclerosis [J]. *J Cell Biol*, 2015, **209**(1): 13-22.
- [5] Cinoku II, Mavragani CP, Moutsopoulos HM. Atherosclerosis: beyond the lipid storage hypothesis. The role of autoimmunity [J]. *Eur J Clin Invest*, 2020, **50**(2): e13195.
- [6] Dai X, Zhang D, Wang C, et al. The pivotal role of thymus in atherosclerosis mediated by immune and inflammatory response [J]. *Int J Med Sci*, 2018, **15**(13): 1555-1563.
- [7] Hassan M. CANTOS: a breakthrough that proves the inflammatory hypothesis of atherosclerosis [J]. *Glob Cardiol Sci Pract*, 2018, **2018**(1): 2.
- [8] Weber C, von Hundelshausen P. CANTOS Trial validates the inflammatory pathogenesis of atherosclerosis setting the stage for a new chapter in therapeutic targeting [J]. *Circ Res*, 2017, **121**(10): 1119-1121.
- [9] Kim JM, Lee WS, Kim J. Therapeutic strategy for atherosclerosis based on bone-vascular axis hypothesis [J]. *Pharmacol Ther*, 2020, **206**: 107436.
- [10] Reis DRA, Medeiros-Fonseca B, Costa JM, et al. HPV infection as a risk factor for atherosclerosis: a connecting hypothesis [J]. *Med Hypotheses*, 2020, **144**: 109979.
- [11] Mai W, Liao Y. Targeting IL-1 beta in the treatment of atherosclerosis [J]. *Front Immunol*, 2020, **11**: 589654.
- [12] Neele AE, Willemsen L, Chen HJ, et al. Targeting epigenetics as atherosclerosis treatment: an updated view [J]. *Curr Opin Lipidol*, 2020, **31**(6): 324-330.
- [13] Wang R, Wang M, Ye J, et al. Mechanism overview and target mining of atherosclerosis: endothelial cell injury in atherosclerosis is regulated by glycolysis (Review) [J]. *Int J Mol Med*, 2021, **47**(1): 65-76.

- [14] Wu WK, Ivanova EA, Orekhov AN. Gut microbiome: a possible common therapeutic target for treatment of atherosclerosis and cancer [J]. *Semin Cancer Biol*, 2021, **70**: 85-97.
- [15] Orekhov AN, Ivanova EA. Cellular models of atherosclerosis and their implication for testing natural substances with anti-atherosclerotic potential [J]. *Phytomedicine*, 2016, **23**(11): 1190-1197.
- [16] Wang D, Yang Y, Lei Y, et al. Targeting foam cell formation in atherosclerosis: therapeutic potential of natural products [J]. *Pharmacol Rev*, 2019, **71**(4): 596-670.
- [17] Zhang S, Li L, Chen W, et al. Natural products: the role and mechanism in low-density lipoprotein oxidation and atherosclerosis [J]. *Phytother Res*, 2021, **35**(6): 2945-2967.
- [18] Li H, Bai L, Qin Q, et al. Research progress on anti-atherosclerosis effect and mechanism of flavonoids compounds mediated by macrophages [J]. *Chin J Chin Mater Med*, 2020, **45**(12): 2827-2834.
- [19] Falk E. Pathogenesis of atherosclerosis [J]. *J Am Coll Cardiol*, 2006, **47**(8): C7-C12.
- [20] Glass CK, Witztum JL. Atherosclerosis: the road ahead [J]. *Cell*, 2001, **104**(4): 503-516.
- [21] Ali AH, Younis N, Abdallah R, et al. Lipid-Lowering therapies for atherosclerosis: statins, fibrates, ezetimibe and PCSK9 monoclonal antibodies [J]. *Curr Med Chem*, 2021, **28**(36): 7427-7445.
- [22] Luis MVJ, Rodrigues-Diez R, Martinez-Lopez D, et al. Oxidative stress in human atherothrombosis: sources, markers and therapeutic targets [J]. *Int J Mol Sci*, 2017, **18**(11): 2315.
- [23] Cayatte AJ, Rupin A, Oliver-Krasinski J, et al. S17834, a new inhibitor of cell adhesion and atherosclerosis that targets NADPH oxidase [J]. *Arterioscler Thromb Vasc Biol*, 2001, **21**(10): 1577-1584.
- [24] Harats D, Shaish A, George J, et al. Overexpression of 15-lipoxygenase in vascular endothelium accelerates early atherosclerosis in LDL receptor-deficient mice [J]. *Arterioscler Thromb Vasc Biol*, 2000, **20**(9): 2100-2105.
- [25] Cyrus T, Witztum JL, Rader DJ, et al. Disruption of the 12/15-lipoxygenase gene diminishes atherosclerosis in apo E-deficient mice [J]. *J Clin Invest*, 1999, **103**(11): 1597-1604.
- [26] Podrez EA, Abu-Soud HM, Hazen SL. Myeloperoxidase-generated oxidants and atherosclerosis [J]. *Free Radic Biol Med*, 2000, **28**(12): 1717-1725.
- [27] Cheng D, Talib J, Stanley CP, et al. Inhibition of MPO (myeloperoxidase) attenuates endothelial dysfunction in mouse models of vascular inflammation and atherosclerosis [J]. *Arterioscler Thromb Vasc Biol*, 2019, **39**(7): 1448-1457.
- [28] Sukhorukov VN, Khotina VA, Chegodaev YS, et al. Lipid metabolism in macrophages: focus on atherosclerosis [J]. *Bio-medicines*, 2020, **8**(8): 262.
- [29] Vazquez MM, Gutierrez MV, Salvatore SR, et al. Nitro-oleic acid, a ligand of CD36, reduces cholesterol accumulation by modulating oxidized-LDL uptake and cholesterol efflux in RAW264. 7 macrophages [J]. *Redox Biol*, 2020, **36**: 101591.
- [30] Cheng XL, Ding F, Wang DP, et al. Hexarelin attenuates atherosclerosis via inhibiting LOX-1-NF-kappa B signaling pathway-mediated macrophage ox-LDL uptake in ApoE^(-/-) mice [J]. *Peptides*, 2019, **121**: 170122.
- [31] Meurs I, Van Eck M, Van Berkel TJC. High-density lipoprotein: key molecule in cholesterol efflux and the prevention of atherosclerosis [J]. *Curr Pharm Des*, 2010, **16**(13): 1445-1467.
- [32] Gou S, Wang L, Zhong C, et al. A novel apoA-I mimetic peptide suppresses atherosclerosis by promoting physiological HDL function in ApoE^(-/-) mice [J]. *Br J Pharmacol*, 2020, **177**(20): 4627-4644.
- [33] Li YY, Zhou SH, Chen SS, et al. PRMT2 inhibits the formation of foam cell induced by ox-LDL in RAW 264. 7 macrophage involving ABCA1 mediated cholesterol efflux [J]. *Biochem Biophys Res Commun*, 2020, **524**(1): 77-82.
- [34] Ma T, Zhang Z, Chen Y, et al. Delivery of nitric oxide in the cardiovascular system: implications for clinical diagnosis and therapy [J]. *Int J Mol Sci*, 2021, **22**(22): 12166. DOI: 10.3390/ijms222212166
- [35] Hong FF, Liang XY, Liu W, et al. Roles of eNOS in atherosclerosis treatment [J]. *Inflamm Res*, 2019, **68**(6): 429-441.
- [36] Balakumar P, Kathuria S, Taneja G, et al. Is targeting eNOS a key mechanistic insight of cardiovascular defensive potentials of statins? [J]. *J Mol Cell Cardiol*, 2012, **52**(1): 83-92.
- [37] Paone S, Baxter AA, Hulett MD, et al. Endothelial cell apoptosis and the role of endothelial cell-derived extracellular vesicles in the progression of atherosclerosis [J]. *Cell Mol Life Sci*, 2019, **76**(6): 1093-1106.
- [38] Ma L, Zheng H, Zhang T. IL-10 suppress vascular smooth muscle cell apoptosis via JAK2/STAT3 signaling pathway and its mechanism of action in atherosclerosis [J]. *Minerva Endocrinol*, 2019, **44**(4): 402-405.
- [39] Qin M, Luo Y, Meng XB, et al. Myricitrin attenuates endothelial cell apoptosis to prevent atherosclerosis: an insight into PI3K/Akt activation and STAT3 signaling pathways [J]. *Vascul Pharmacol*, 2015, **70**: 23-34.
- [40] Zhong X, Ma X, Zhang L, et al. MIAT promotes proliferation and hinders apoptosis by modulating miR-181b/STAT3 axis in ox-LDL-induced atherosclerosis cell models [J]. *Biomed Pharmacother*, 2018, **97**: 1078-1085.
- [41] Xiong X, Lu W, Zhang K, et al. Pterostilbene reduces endothelial cell apoptosis by regulation of the Nrf2-mediated TLR-4/MyD88/NF-kappa B pathway in a rat model of atherosclerosis [J]. *Exp Ther Med*, 2020, **20**(3): 2090-2098.
- [42] Yang L, Gao C. MiR-590 inhibits endothelial cell apoptosis by inactivating the TLR4/NF-kappa B pathway in atherosclerosis [J]. *Yonsei Med J*, 2019, **60**(3): 298-307.
- [43] Xu K, Liu X, Yin D, et al. PP2A alleviates oxidized LDL-induced endothelial dysfunction by regulating LOX-1/ROS/MAPK axis [J]. *Life Sci*, 2020, **243**: 117270.
- [44] Geng J, Fu W, Yu X, et al. Ginsenoside Rg3 alleviates ox-LDL induced endothelial dysfunction and prevents atherosclerosis in ApoE^(-/-) mice by regulating PPAR gamma/FAK signaling pathway [J]. *Front Pharmacol*, 2020, **11**: 500.
- [45] Langbein H, Brunssen C, Hofmann A, et al. NADPH oxidase 4 protects against development of endothelial dysfunction and atherosclerosis in LDL receptor deficient mice [J]. *Eur Heart J*, 2016, **37**(22): 1753-1761.
- [46] Tedgui A, Mallat Z. Cytokines in atherosclerosis: pathogenic and regulatory pathways [J]. *Physiol Rev*, 2006, **86**(2): 515-581.
- [47] Gora IM, Ciechanowska A, Ladyzynski P. NLRP3 inflammasome at the interface of inflammation, endothelial dysfunction, and type 2 diabetes [J]. *Cells*, 2021, **10**(2): 314.
- [48] Li W, Cao T, Luo C, et al. Crosstalk between ER stress, NLRP3 inflammasome, and inflammation [J]. *Appl Microbiol Biotechnol*, 2020, **104**(14): 6129-6140.
- [49] Bhaskar V, Yin J, Mirza AM, et al. Monoclonal antibodies targeting IL-1 beta reduce biomarkers of atherosclerosis *in vitro* and inhibit atherosclerotic plaque formation in Apolipoprotein E-deficient mice [J]. *Atherosclerosis*, 2011, **216**(2): 313-320.
- [50] Tracey D, Klareskog L, Sasso EH, et al. Tumor necrosis factor antagonist mechanisms of action: a comprehensive review [J]. *Pharmacol Ther*, 2008, **117**(2): 244-279.
- [51] Saigusa R, Winkels H, Ley K. T cell subsets and functions in

- atherosclerosis [J]. *Nat Rev Cardiol*, 2020, **17**(7): 387-401.
- [52] Marchini T, Hansen S, Wolf D. ApoB-specific CD4⁺ T cells in mouse and human atherosclerosis [J]. *Cells*, 2021, **10**(2): 446.
- [53] Schaefer S, Zernecke A. CD8(+) T cells in atherosclerosis [J]. *Cells*, 2021, **10**(1): 37.
- [54] Poels K, van Leent MMT, Boutros C, et al. Immune checkpoint inhibitor therapy aggravates T cell-driven plaque inflammation in atherosclerosis [J]. *JACC CardioOncol*, 2020, **2**(4): 599-610.
- [55] Zhao TX, Kostapanos M, Griffiths C, et al. Low-dose interleukin-2 in patients with stable ischaemic heart disease and acute coronary syndromes (LILACS): protocol and study rationale for a randomised, double-blind, placebo-controlled, phase I/II clinical trial [J]. *BMJ Open*, 2018, **8**(9): e022452.
- [56] Schrijvers DM, De Meyer GRY, Martinet W. Autophagy in atherosclerosis a potential drug target for plaque stabilization [J]. *Arterioscler Thromb Vasc Biol*, 2011, **31**(12): 2787-2791.
- [57] Ou H, Liu C, Feng W, et al. Role of AMPK in atherosclerosis via autophagy regulation [J]. *Sci Chin Life Sci*, 2018, **61**(10): 1212-1221.
- [58] Testa G, Staurengi E, Giannelli S, et al. Up-regulation of PC-SK6 by lipid oxidation products: a possible role in atherosclerosis [J]. *Biochimie*, 2021, **181**: 191-203.
- [59] Jialal I, Fuller CJ. Effect of vitamin E, vitamin C and beta-carotene on LDL oxidation and atherosclerosis [J]. *Can J Cardiol*, 1995, **11**: G97-G103.
- [60] Reaven PD, Khouw A, Beltz WF, et al. Effect of dietary antioxidant combinations in humans. Protection of LDL by vitamin E but not by beta-carotene [J]. *Arterioscler Thromb*, 1993, **13**(4): 590-600.
- [61] Ganini D, Mason RP. Absence of an effect of vitamin E on protein and lipid radical formation during lipoperoxidation of LDL by lipoxygenase [J]. *Free Radic Biol Med*, 2014, **76**: 61-68.
- [62] Moser MA, Chun OK. Vitamin C and heart health: a review based on findings from epidemiologic studies [J]. *Int J Mol Sci*, 2016, **17**(8): 1328.
- [63] Jafarpur M, Jafarpur S. Protective effects of vitamin C on atherosclerosis induced by hypervitaminosis D [J]. *Int J Cardiol*, 2011, **147**: S13-S14.
- [64] Das S, Snehlata, Srivastava LM. Antioxidant effect of vitamin C in hypercholesterolemic atherosclerosis [J]. *FASEB J*, 1997, **11**(9): A1432-A1432.
- [65] Babaev VR, Li L, Shah S, et al. Combined vitamin C and vitamin E deficiency worsens early atherosclerosis in apolipoprotein E-deficient mice [J]. *Arterioscler Thromb Vasc Biol*, 2010, **30**(9): 1751-1757.
- [66] Goralska J, Hartwich J, Siedlecka D, et al. Antioxidative effect of combined vitamin E and C supplementation in men with risk of atherosclerosis [J]. *Atheroscler Suppl*, 2007, **8**(1): 169-169.
- [67] Ekuni D, Tomofuji T, Sanbe T, et al. Vitamin C intake attenuates the degree of experimental atherosclerosis induced by periodontitis in the rat by decreasing oxidative stress [J]. *Arch Oral Biol*, 2009, **54**(5): 495-502.
- [68] Xiao L, Liu L, Guo X, et al. Quercetin attenuates high fat diet-induced atherosclerosis in apolipoprotein E knockout mice: a critical role of NADPH oxidase [J]. *Food Chem Toxicol*, 2017, **105**: 22-33.
- [69] Aviram M. HDL-associated paraoxonase 1 (PON1) and dietary antioxidants attenuate upoprotein oxidation, macrophage foam cells formation and atherosclerosis development [J]. *Pathophysiol Haemost Thromb*, 2006, **35**(1-2): 146-151.
- [70] Guseva DA, Khudoklinova YY, Medvedeva NV, et al. Influence of resveratrol and dihydroquercetin inclusion into phospholipid nanopatrics on their bioavailability and specific activity [J]. *Biomed Khim*, 2015, **61**(5): 598-605.
- [71] Miura Y, Chiba T, Tomita I, et al. Tea catechins prevent the development of atherosclerosis in apoprotein E-deficient mice [J]. *J Nutr*, 2001, **131**(1): 27-32.
- [72] Thuong PT, Pokharel YR, Lee MY, et al. Dual anti-oxidative effects of fraxetin isolated from *Fraxinus rhynchophylla* [J]. *Biol Pharm Bull*, 2009, **32**(9): 1527-1532.
- [73] Feng Y, Wu J, Cong R, et al. The effect of neferine on foam cell formation by anti-low density lipoprotein oxidation [J]. *J Tongji Med Univ*, 1998, **18**(3): 134-136.
- [74] Lian N, Tong J, Li W, et al. Ginkgetin ameliorates experimental atherosclerosis in rats [J]. *Biomed Pharmacother*, 2018, **102**: 510-516.
- [75] Lu Y, He Z, Shen X, et al. Cholesterol-lowering effect of allixin on hypercholesterolemic ICR mice [J]. *Oxid Med Cell Longev*, 2012, **2012**: 489690.
- [76] Zhang WP, Bai XJ, Zheng XP, et al. Icarin attenuates the enhanced prothrombotic state in atherosclerotic rabbits independently of its lipid-lowering effects [J]. *Planta Med*, 2013, **79**(9): 731-736.
- [77] Hirata H, Yimin, Segawa S, et al. Xanthohumol prevents atherosclerosis by reducing arterial cholesterol content via CETP and apolipoprotein E in CETP-transgenic mice [J]. *PLoS One*, 2012, **7**(11): e49415.
- [78] Beg M, Singhal KC, Afzaal S. A study of effect of guggulsterone on hyperlipidemia of secondary glomerulopathy [J]. *Indian J Physiol Pharmacol*, 1996, **40**(3): 237-240.
- [79] Ellinghaus P, Wolfrum C, Assmann G, et al. Phytanic acid activates the peroxisome proliferator-activated receptor alpha (PPAR alpha) in sterol carrier protein 2- sterol carrier protein x-deficient mice [J]. *J Biol Chem*, 1999, **274**(5): 2766-2772.
- [80] Narender T, Shweta S, Tiwari P, et al. Antihyperglycemic and antidiabetic agent from *Aegle marmelos* [J]. *Bioorg Med Chem Lett*, 2007, **17**(6): 1808-1811.
- [81] Singh SP, Sashidhara KV. Lipid lowering agents of natural origin: an account of some promising chemotypes [J]. *Eur J Med Chem*, 2017, **140**: 331-348.
- [82] Tang FT, Cao Y, Wang TQ, et al. Tanshinone IIA attenuates atherosclerosis in ApoE(-/-) mice through down-regulation of scavenger receptor expression [J]. *Eur J Pharmacol*, 2011, **650**(1): 275-284.
- [83] Ricciarelli R, Zingg JM, Azzi A. Vitamin E reduces the uptake of oxidized LDL by inhibiting CD36 scavenger receptor expression in cultured aortic smooth muscle cells [J]. *Circulation*, 2000, **102**(1): 82-87.
- [84] Wang S, Zhang X, Liu M, et al. Chrysin inhibits foam cell formation through promoting cholesterol efflux from RAW264. 7 macrophages [J]. *Pharm Biol*, 2015, **53**(10): 1481-1487.
- [85] Zhou Y, Chen R, Liu D, et al. Asperlin inhibits LPS-evoked foam cell formation and prevents atherosclerosis in ApoE^{-/-} mice [J]. *Mar Drugs*, 2017, **15**(11): 358.
- [86] Lu S, Luo Y, Sun G, et al. Ginsenoside compound K attenuates ox-LDL-mediated macrophage inflammation and foam cell formation via autophagy induction and modulating NF-kappa B, p38, and JNK MAPK signaling [J]. *Front Pharmacol*, 2020, **11**: 567238.
- [87] Xu Y, Liu Q, Xu Y, et al. Rutaecarpine suppresses atherosclerosis in ApoE^{-/-} mice through upregulating ABCA1 and SR-BI within RCT [J]. *J Lipid Res*, 2014, **55**(8): 1634-1647.
- [88] Jiang Z, Sang H, Fu X, et al. Alpinetin enhances cholesterol efflux and inhibits lipid accumulation in oxidized low-density

- lipoprotein-loaded human macrophages [J]. *Biotechnol Appl Biochem*, 2015, **62**(6): 840-847.
- [89] He XW, Yu D, Li WL, et al. Anti-atherosclerotic potential of baicalin mediated by promoting cholesterol efflux from macrophages via the PPAR gamma-LXR alpha-ABCA1/ABCG1 pathway [J]. *Biomed Pharmacother*, 2016, **83**: 257-264.
- [90] Palozza P, Simone R, Catalano A, et al. Lycopene regulation of cholesterol synthesis and efflux in human macrophages [J]. *J Nutr Biochem*, 2011, **22**(10): 971-978.
- [91] Zhao W, Haller V, Ritsch A. The polyphenol PGG enhances expression of SR-BI and ABCA1 in J774 and THP-1 macrophages [J]. *Atherosclerosis*, 2015, **242**(2): 611-617.
- [92] Wang D, Hiebl V, Ladurner A, et al. 6-Dihydroparadol, a ginger constituent, enhances cholesterol efflux from THP-1-derived macrophages [J]. *Mol Nutr Food Res*, 2018, **62**(14): e1800011.
- [93] Li X, Zhou Y, Yu C, et al. Paeonol suppresses lipid accumulation in macrophages via upregulation of the ATP-binding cassette transporter A1 and downregulation of the cluster of differentiation 36 [J]. *Int J Oncol*, 2015, **46**(2): 764-774.
- [94] Xu X, Li Q, Pang L, et al. Arctigenin promotes cholesterol efflux from THP-1 macrophages through PPAR-gamma/LXR-alpha signaling pathway [J]. *Biochem Biophys Res Commun*, 2013, **441**(2): 321-326.
- [95] Park SH, Paek JH, Shin D, et al. Purple perilla extracts with alpha-asarone enhance cholesterol efflux from oxidized LDL-exposed macrophages [J]. *Int J Mol Med*, 2015, **35**(4): 957-965.
- [96] Wu C, Luan H, Zhang X, et al. Chlorogenic acid protects against atherosclerosis in ApoE^{-/-} mice and promotes cholesterol efflux from RAW264. 7 macrophages [J]. *PLoS One*, 2014, **9**(9): e95452.
- [97] Chi L, Peng L, Hu X, et al. Berberine combined with atorvastatin downregulates LOX-1 expression through the ET-1 receptor in monocyte/macrophages [J]. *Int J Mol Med*, 2014, **34**(1): 283-290.
- [98] Wang L, Palme V, Rotter S, et al. Piperine inhibits ABCA1 degradation and promotes cholesterol efflux from THP-1-derived macrophages [J]. *Mol Nutr Food Res*, 2017, **61**(4): 1500960.
- [99] Jiang T, Ren K, Chen Q, et al. Leonurine prevents atherosclerosis via promoting the expression of ABCA1 and ABCG1 in a Ppar gamma/Lxr alpha signaling pathway-dependent manner [J]. *Cell Physiol Biochem*, 2017, **43**(4): 1703-1717.
- [100] Ricci A, Iaccheri E, Benelli A, et al. Rapid optical method for procyanidins estimation in red wines [J]. *Food Control*, 2020, **118**: 107439.
- [101] Wang K, Chen X, Chen Y, et al. Grape seed procyanidins suppress the apoptosis and senescence of chondrocytes and ameliorates osteoarthritis via the DPP4-Sirt1 pathway [J]. *Food Funct*, 2020, **11**(12): 10493-10505.
- [102] Yang H, Xiao L, Yuan Y, et al. Procyanidin B2 inhibits NLRP3 inflammasome activation in human vascular endothelial cells [J]. *Biochem Pharmacol*, 2014, **92**(4): 599-606.
- [103] Zhang X, Wang G, Gurley EC, et al. Flavonoid apigenin inhibits lipopolysaccharide-induced inflammatory response through multiple mechanisms in macrophages [J]. *PLoS One*, 2014, **9**(9): e107072.
- [104] Hu Q, Zhang T, Yi L, et al. Dihydromyricetin inhibits NLRP3 inflammasome-dependent pyroptosis by activating the Nrf2 signaling pathway in vascular endothelial cells [J]. *Biofactors*, 2018, **44**(2): 123-136.
- [105] Zhang J, Yan J. Protective effect of ginkgolic acid in attenuating LDL induced inflammation human peripheral blood mononuclear cells via altering the NF-kappa B signaling pathway [J]. *Front Pharmacol*, 2019, **10**: 1241.
- [106] Yu J, Ming H, Li HY, et al. IMM-H007, a novel small molecule inhibitor for atherosclerosis, represses endothelium inflammation by regulating the activity of NF-kappa B and JNK/AP1 signaling [J]. *Toxicol Appl Pharmacol*, 2019, **381**: 114732.
- [107] Zhang X, Xue C, Xu Q, et al. Caprylic acid suppresses inflammation via TLR4/NF-kappa B signaling and improves atherosclerosis in ApoE-deficient mice [J]. *Nutr Metab (Lond)*, 2019, **16**: 40.
- [108] Choi JH, Jeong TS, Kim DY, et al. Hematein inhibits atherosclerosis by inhibition of reactive oxygen generation and NF-kappa B-dependent inflammatory mediators in hyperlipidemic mice [J]. *J Cardiovasc Pharmacol*, 2003, **42**(2): 287-295.
- [109] Srivastava RAK, Mistry S, Sharma S. A novel anti-inflammatory natural product from *Sphaeranthus indicus* inhibits expression of VCAM1 and ICAM1, and slows atherosclerosis progression independent of lipid changes [J]. *Nutr Metab (Lond)*, 2015, **12**: 20.
- [110] Bhaskar S, Sudhakaran PR, Helen A. Quercetin attenuates atherosclerotic inflammation and adhesion molecule expression by modulating TLR-NF-kappa B signaling pathway [J]. *Cell Immunol*, 2016, **310**: 131-140.
- [111] Panicker SR, Sreenivas P, Babu MS, et al. Quercetin attenuates monocyte chemoattractant protein-1 gene expression in glucose primed aortic endothelial cells through NF-kappa B and AP-1 [J]. *Pharmacol Res*, 2010, **62**(4): 328-336.
- [112] Zhang F, Feng J, Zhang J, et al. Quercetin modulates AMPK/SIRT1/NF-kappa B signaling to inhibit inflammatory/oxidative stress responses in diabetic high fat diet-induced atherosclerosis in the rat carotid artery [J]. *Exp Ther Med*, 2020, **20**(6): 280.
- [113] Zhao J, Quan X, Xie Z, et al. Juglanin suppresses oscillatory shear stress-induced endothelial dysfunction: an implication in atherosclerosis [J]. *Int Immunopharmacol*, 2020, **89**: 107048.
- [114] Elseweidy MM, Amin RS, Atteia HH, et al. New insight on a combination of policosanol and 10-dehydrogingerdione phytochemicals as inhibitors for platelet activation biomarkers and atherogenicity risk in dyslipidemic rabbits: role of CETP and PCSK9 inhibition [J]. *Appl Biochem Biotechnol*, 2018, **186**(4): 805-815.
- [115] Lü SL, Dang GH, Deng JC, et al. Shikonin attenuates hyperhomocysteinemia-induced CD4(+) T cell inflammatory activation and atherosclerosis in ApoE^{-/-} mice by metabolic suppression [J]. *Acta Pharmacol Sin*, 2020, **41**(1): 47-55.
- [116] Devi KP, Malar DS, Nabavi SF, et al. Kaempferol and inflammation: from chemistry to medicine [J]. *Pharmacol Res*, 2015, **99**: 1-10.
- [117] Tabares-Guevara JH, Lara-Guzman OJ, Londono-Londono JA, et al. Natural B flavonoids modulate macrophage-oxidized LDL interaction in vitro and promote atheroprotection in vivo [J]. *Front Immunol*, 2017, **8**: 923.
- [118] Wu FC, Jiang JG. Effects of diosgenin and its derivatives on atherosclerosis [J]. *Food Funct*, 2019, **10**(11): 7022-7036.
- [119] Voloshyna I, Hussaini SM, Reiss AB. Resveratrol in cholesterol metabolism and atherosclerosis [J]. *J Med Food*, 2012, **15**(9): 763-773.
- [120] Lee RT, Libby P. The unstable atheroma [J]. *Arterioscler Thromb Vasc Biol*, 1997, **17**(10): 1859-1867.
- [121] Tian K, Ogura S, Little PJ, et al. Targeting LOX-1 in atherosclerosis and vasculopathy: current knowledge and future perspectives [J]. *Ann N Y Acad Sci*, 2019, **1443**(1): 34-53.
- [122] Liu Z, Xu S, Huang X, et al. Cryptotanshinone, an orally bioactive herbal compound from Danshen, attenuates atherosclerosis in apolipoprotein E-deficient mice: role of lectin-like ox-

- idized LDL receptor-1 (LOX-1) [J]. *Br J Pharmacol*, 2015, **172**(23): 5661-5675.
- [123] Guan S, Wang B, Li W, *et al.* Effects of berberine on expression of LOX-1 and SR-BI in human macrophage-derived foam cells induced by ox-LDL [J]. *Am J Chin Med*, 2010, **38**(6): 1161-1169.
- [124] Qin M, Luo Y, Lu S, *et al.* Ginsenoside F1 ameliorates endothelial cell inflammatory injury and prevents atherosclerosis in mice through A20-mediated suppression of NF-kappa B signaling [J]. *Front Pharmacol*, 2017, **8**: 953.
- [125] Siti HN, Kamisah Y, Kamsiah J. The role of oxidative stress, antioxidants and vascular inflammation in cardiovascular disease (a review) [J]. *Vascul Pharmacol*, 2015, **71**: 40-56.
- [126] Zhang C, Wang C, Li W, *et al.* Pharmacokinetics and pharmacodynamics of the triterpenoid ursolic acid in regulating the antioxidant, anti-inflammatory, and epigenetic gene responses in rat leukocytes [J]. *Mol Pharm*, 2017, **14**(11): 3709-3717.
- [127] Li Q, Zhao W, Zeng X, *et al.* Ursolic acid attenuates atherosclerosis in ApoE^{-/-} mice: role of LOX-1 mediated by ROS/NF-B pathway [J]. *Molecules*, 2018, **23**(5): 1101.
- [128] Zhang Z, Zhang D, Du B, *et al.* Hyperoside inhibits the effects induced by oxidized low-density lipoprotein in vascular smooth muscle cells via oxLDL-LOX-1-ERK pathway [J]. *Mol Cell Biochem*, 2017, **433**(1-2): 169-176.
- [129] Gu L, Bai W, Li S, *et al.* Celastrol prevents atherosclerosis via inhibiting LOX-1 and oxidative stress [J]. *PLoS One*, 2013, **8**(6): e65477.
- [130] Zhang L, Jia YH, Zhao XS, *et al.* Trichosanatine alleviates oxidized low-density lipoprotein induced endothelial cells injury via inhibiting the LOX-1/p38 MAPK pathway [J]. *Am J Transl Res*, 2016, **8**(12): 5455-5464.
- [131] Zhao W, Li C, Gao H, *et al.* Dihydrotanshinone I attenuates atherosclerosis in ApoE-deficient mice: role of NOX4/NF-kappa B mediated lectin-like oxidized LDL receptor-1 (LOX-1) of the endothelium [J]. *Front Pharmacol*, 2016, **7**: 418.
- [132] Teng CM, Hsueh CM, Chang YL, *et al.* Antiplatelet effects of some aporphine and phenanthrene alkaloids in rabbits and man [J]. *J Pharm Pharmacol*, 1997, **49**(7): 706-711.
- [133] Song N, Jia L, Cao H, *et al.* Gypenoside inhibits endothelial cell apoptosis in atherosclerosis by modulating mitochondria through PI3K/Akt/Bad pathway [J]. *Biomed Res Int*, 2020, **2020**: 2819658.
- [134] Han WM, Chen XC, Li GR, *et al.* Acacetin protects against high glucose-induced endothelial cells injury by preserving mitochondrial function via activating Sirt1/Sirt3/AMPK signals [J]. *Front Pharmacol*, 2020, **11**: 607796.
- [135] Sun GB, Qin M, Ye JX, *et al.* Inhibitory effects of myricitrin on oxidative stress-induced endothelial damage and early atherosclerosis in ApoE^{-/-} mice [J]. *Toxicol Appl Pharmacol*, 2013, **271**(1): 114-126.
- [136] Luo S, Li H, Mo Z, *et al.* Connectivity map identifies luteolin as a treatment option of ischemic stroke by inhibiting MMP9 and activation of the PI3K/Akt signaling pathway [J]. *Exp Mol Med*, 2019, **51**(3): 1-11.
- [137] Luo Y, Sun G, Dong X, *et al.* Isorhamnetin attenuates atherosclerosis by inhibiting macrophage apoptosis via PI3K/AKT activation and HO-1 induction [J]. *PLoS One*, 2015, **10**(3): e0120259.
- [138] Zhang S, Guo C, Chen Z, *et al.* Vitexin alleviates ox-LDL-mediated endothelial injury by inducing autophagy via AMPK signaling activation [J]. *Mol Immunol*, 2017, **85**: 214-221.
- [139] Saha S, Profumo E, Togna AR, *et al.* Lupeol counteracts the proinflammatory signalling triggered by 7-Keto-cholesterol: new perspectives in the therapy of atherosclerosis [J]. *Oxid Med Cell Longev*, 2020, **2020**: 1232816.
- [140] Guo S, Long M, Li X, *et al.* Curcumin activates autophagy and attenuates oxidative damage in EA. hy926 cells via the Akt/mTOR pathway [J]. *Mol Med Report*, 2016, **13**(3): 2187-2193.
- [141] Wang K, Liu R, Li J, *et al.* Quercetin induces protective autophagy in gastric cancer cells involvement of Akt-mTOR- and hypoxia-induced factor 1 alpha-mediated signaling [J]. *Autophagy*, 2011, **7**(9): 966-978.
- [142] Lee GH, Park JS, Jin SW, *et al.* Betulinic acid induces eNOS expression via the AMPK-dependent KLF2 signaling pathway [J]. *J Agric Food Chem*, 2020, **68**(49): 14523-14530.
- [143] Lin CP, Huang PH, Tsai HS, *et al.* Monascus purpureus-fermented rice inhibits tumor necrosis factor-alpha-induced up-regulation of matrix metalloproteinase 2 and 9 in human aortic smooth muscle cells [J]. *J Pharm Pharmacol*, 2011, **63**(12): 1587-1594.
- [144] Dong M, Zhou C, Ji L, *et al.* AG1296 enhances plaque stability via inhibiting inflammatory responses and decreasing MMP-2 and MMP-9 expression in ApoE^{-/-} mice [J]. *Biochem Biophys Res Commun*, 2017, **489**(4): 426-431.
- [145] Li W, Niu X, Yu J, *et al.* Imperatorin alleviates the abnormal proliferation, migration, and foaming of ox-LDL-induced VSMCs through regulating PI3K/Akt/mTOR signaling pathway [J]. *J Funct Foods*, 2020, **70**: 103982.
- [146] Lee YJ, Han BH, Yoon JJ, *et al.* Identification of securinine as vascular protective agent targeting atherosclerosis in vascular endothelial cells, smooth muscle cells, and apolipoprotein E deficient mice [J]. *Phytomedicine*, 2021, **81**: 153430.
- [147] Guo M, Guo G, Xiao J, *et al.* Ginsenoside Rg3 stereoisomers differentially inhibit vascular smooth muscle cell proliferation and migration in diabetic atherosclerosis [J]. *J Cell Mol Med*, 2018, **22**(6): 3202-3214.
- [148] Wang Y, Huang ZQ, Wang CQ, *et al.* Artemisinin inhibits extracellular matrix metalloproteinase inducer (EMMPRIN) and matrix metalloproteinase-9 expression via a protein kinase C delta/p38/extracellular signal-regulated kinase pathway in phorbol myristate acetate-induced THP-1 macrophages [J]. *Clin Exp Pharmacol Physiol*, 2011, **38**(1): 11-18.
- [149] Paper DH. Natural products as angiogenesis inhibitors [J]. *Planta Med*, 1998, **64**(8): 686-695.
- [150] Morrison DJ, Preston T. Formation of short chain fatty acids by the gut microbiota and their impact on human metabolism [J]. *Gut Microbes*, 2016, **7**(3): 189-200.
- [151] Zhao X, Oduro PK, Tong W, *et al.* Therapeutic potential of natural products against atherosclerosis: targeting on gut microbiota [J]. *Pharmacol Res*, 2021, **163**: 105362.
- [152] Duttaray AK. Role of gut microbiota and their metabolites on atherosclerosis, hypertension and human blood platelet function: a review [J]. *Nutrients*, 2021, **13**(1): 144.
- [153] Chen PB, Black AS, Sobel AL, *et al.* Directed remodeling of the mouse gut microbiome inhibits the development of atherosclerosis [J]. *Nat Biotechnol*, 2020, **38**(11): 1288-1297.
- [154] Feng X, Sureda A, Jafari S, *et al.* Berberine in cardiovascular and metabolic diseases: from mechanisms to therapeutics [J]. *Theranostics*, 2019, **9**(7): 1923-1951.
- [155] Zhu L, Zhang D, Zhu H, *et al.* Berberine treatment increases Akkermansia in the gut and improves high-fat diet-induced atherosclerosis in ApoE^{-/-} mice [J]. *Atherosclerosis*, 2018, **268**: 117-126.
- [156] Wu M, Yang S, Wang S, *et al.* Effect of berberine on atherosclerosis and gut microbiota modulation and their correlation in high-fat diet-fed ApoE^{-/-} mice [J]. *Front Pharmacol*, 2020, **11**: 223.
- [157] Ma Y, Chen K, Lv L, *et al.* Ferulic acid ameliorates nonalco-

- holic fatty liver disease and modulates the gut microbiota composition in high-fat diet fed ApoE^{-/-} mice [J]. *Biomed Pharmacother*, 2019, **113**: 108753.
- [158] Neyrinck AM, Catry E, Taminiau B, *et al.* Chitin-glucan and pomegranate polyphenols improve endothelial dysfunction [J]. *Sci Rep*, 2019, **9**(1): 14150.
- [159] Wang F, Zhao C, Tian G, *et al.* Naringin alleviates atherosclerosis in ApoE^{-/-} mice by regulating cholesterol metabolism involved in gut microbiota remodeling [J]. *J Agric Food Chem*, 2020, **68**(45): 12651-12660.
- [160] Luo Y, Fang JL, Yuan K, *et al.* Ameliorative effect of purified anthocyanin from *Lycium ruthenicum* on atherosclerosis in rats through synergistic modulation of the gut microbiota and NF-kappa B/SREBP-2 pathways [J]. *J Funct Foods*, 2019, **59**: 223-233.
- [161] Anwar S, Bhandari U, Panda BP, *et al.* Trigonelline inhibits intestinal microbial metabolism of choline and its associated cardiovascular risk [J]. *J Pharm Biomed Anal*, 2018, **159**: 100-112.
- [162] Li F, Zhang T, He Y, *et al.* Inflammation inhibition and gut microbiota regulation by TSG to combat atherosclerosis in ApoE^{-/-} mice [J]. *J Ethnopharmacol*, 2020, **247**: 112232.
- [163] Lv Z, Shan X, Tu Q, *et al.* Ginkgolide B treatment regulated intestinal flora to improve high-fat diet induced atherosclerosis in ApoE^{-/-} mice [J]. *Biomed Pharmacother*, 2021, **134**: 111100.
- [164] Wilson B, Whelan K. Prebiotic inulin-type fructans and galacto-oligosaccharides: definition, specificity, function, and application in gastrointestinal disorders [J]. *J Gastroenterol Hepatol*, 2017, **32**: 64-68.
- [165] Hoving LR, Katiraei S, Heijink M, *et al.* Dietary mannan oligosaccharides modulate gut microbiota, increase fecal bile acid excretion, and decrease plasma cholesterol and atherosclerosis development [J]. *Mol Nutr Food Res*, 2018, **62**(10): e1700942.
- [166] Pang B, Yue H, Wang E, *et al.* A metabonomics study of atherosclerosis by rapid resolution liquid chromatography quadrupole time-of-flight mass spectrometry [J]. *Chin J Anal Chem*, 2015, **43**(11): 1766-1771.
- [167] Jiang CY, Yang KM, Yang L, *et al.* A ¹H NMR based metabonomics approach to progression of coronary atherosclerosis in a hamster model [J]. *Acta Pharm Sin*, 2013, **48**(4): 495-502.
- [168] Peng J B, Jia H M, Xu T, *et al.* A ¹H NMR based metabonomics approach to progression of coronary atherosclerosis in a rabbit model [J]. *Process Biochem*, 2011, **46**(12): 2240-2247.
- [169] Mourino-Alvarez L, Baldan-Martin M, Rincon R, *et al.* Recent advances and clinical insights into the use of proteomics in the study of atherosclerosis [J]. *Expert Rev Proteomics*, 2017, **14**(8): 701-713.
- [170] Duan H, Khan GJ, Shang LJ, *et al.* Computational pharmacology and bioinformatics to explore the potential mechanism of Schisandra against atherosclerosis [J]. *Food Chem Toxicol*, 2021, **150**: 112058.
- [171] Pan Y, Yu C, Huang J, *et al.* Bioinformatics analysis of vascular RNA-seq data revealed hub genes and pathways in a novel Tibetan minipig atherosclerosis model induced by a high fat/cholesterol diet [J]. *Lipids Health Dis*, 2020, **19**(1): 54.
- [172] Hardung D, Behne A, Langhoff R. Dual Pathway Inhibition in Atherosclerosis-Which Patients Benefit? [J]. *Dtsch Med Wochenschr*, 2019, **144**(20): 1384-1389.
- [173] Wil DN, Guan L, Jiang YX, *et al.* Microbiome and metabonomics study of Quercetin for the treatment of atherosclerosis [J]. *Cardiovasc Diagn Ther*, 2019, **9**(6): 545-560.
- [174] Groner J, Goepferich A, Breunig M. Atherosclerosis: conventional intake of cardiovascular drugs versus delivery using nanotechnology-a new chance for causative therapy? [J]. *J Control Release*, 2021, **333**: 536-559.
- [175] Song Y, Zhang N, Li Q, *et al.* Biomimetic liposomes hybrid with platelet membranes for targeted therapy of atherosclerosis [J]. *Chem Eng J*, 2021, **408**: 127296.
- [176] Zang X, Cheng M, Zhang X, *et al.* Targeting macrophages using nanoparticles: a potential therapeutic strategy for atherosclerosis [J]. *J Mater Chem B*, 2021, **9**(15): 3284-3294.

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